

POSTHARVEST BEHAVIOR OF Highbush BLUEBERRY FRUITS CV. O'NEAL CULTIVATED WITH DIFFERENT ORGANIC FERTILIZATION TREATMENTS

Graciela Echeverría V.¹, Juan Cañumir V.^{2*}, and Humberto Serri G.¹

ABSTRACT

Considering the increasing demand for organic products and the fact that Chile is far from export markets, fresh fruits of highbush blueberry (*Vaccinium corymbosum* L. x *Vaccinium darrowii* Camp) from organic fertilization were stored at 3 °C and 90% RH for 30 d in modified atmosphere (MA) and conventional atmosphere (CA) with the objective of studying the effect of organic fertilization treatments in postharvest behavior. The variables, water content, weight loss, diameter loss, soluble solids, titratable acidity, pH, and presence of pathogens were evaluated during three periods; with a divided plot factorial arrangement experimental design. Results were submitted to variance analysis, and Tukey test ($P \leq 0.05$) was applied when significant differences appeared. Most of the evaluated variables showed no differences between fruits treated with organic or conventional fertilization. Nevertheless, the greater presence of pathogens occurred in fruits treated with organic fertilization when they were stored in CA, being *Botrytis cinerea* the causal agent with greater incidence. The fruits stored in MA presented a better postharvest behavior than the fruits stored in CA, demonstrating less weight loss (2.3%), diameter loss (15.6%), incidence of microorganisms, less variation of soluble solids over time (14.6%), acidity (0.37% citric acid), and humidity (81%).

Key words: organic fertilization, modified atmosphere, *Botrytis*, *Vaccinium corymbosum*.

INTRODUCTION

World blueberry consumption has risen mainly because of its health benefits (Sinelli *et al.*, 2008), and organically produced blueberries are increasingly being consumed. Member countries of the European Union, as well as Japan, demonstrate an ever-increasing demand for organic products (Granatstein, 2000; Shea, 2004; Smith and Marsden, 2004; Sawyer *et al.*, 2008). Almost all organic products are overpriced in relation to conventional products (ProChile, 2003; Kuepper and Diver, 2004).

The majority of blueberry orchards in Chile are conventionally managed with defined management guidelines (Hepp, 2005). However, management of organic blueberry orchards has not yet been defined. There are actually many questions about organic fertilization management because of the scarcity of

supplies that comply with organic requirements (INN, 2004) and its efficiency in terms of production. At the same time, fertilization is one of the principal problems that producers face when changing from conventional to organic production (Morales, 2004). The basis of soil fertility management in organic systems consists in incorporating important quantities of organic matter (OM) through the application of animal and plant origin materials that permit improving soil characteristics. The organic production system does not try to substitute the crop nutrient requirements with soluble fertilizers that inhibit microorganism activity, but aims to increment soil fertility and maintain it in the long-term. However, faced with nutritional deficiencies, it is possible to use several fertilizers of organic and mineral origin as a supplement (Céspedes *et al.*, 2005). The fruits evaluated in this study came from plants treated with different sources of organic fertilization (Table 1) which are applied by organic blueberry producers in Chile.

Organic fertilization permits a balanced nutrient contribution; as a result, it is expected to obtain fruits with a similar or better quality than that produced with conventional fertilization.

Productive efforts to obtain quality fruit can be in

¹Universidad de Concepción, Facultad de Agronomía, Av. Vicente Méndez 595, Casilla 537, Chillán, Chile.

²Universidad de Concepción, Facultad de Ingeniería Agrícola, Av. Vicente Méndez 595, Casilla 537, Chillán, Chile. *Corresponding author (jcanumir@udec.cl).

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vain if after harvest the fruit is not duly handled. This consideration is especially important for Chile since it is far from the markets where it exports fresh fruit, and which requires conservation in a bioclimatic chamber, as well as other techniques that permit maintaining quality. In postharvest the fruits respire at the expense of their nutritive reserves and transpire their own water content if there is no compensation of the respired substrates, initiating deterioration (Loyola *et al.*, 1996; Romojaró *et al.*, 1996; Wills *et al.*, 1998; Cañumir and Loyola, 2004). Modified atmosphere is a fruit conservation method that reduces respiration rate and water loss of the product which implies the replacement of air in a container with a mixture of different gases, but without exerting subsequent control during storage. Usually, the concentration of CO₂ and/or N is increased and O₂ decreased (Romojaró *et al.*, 1996; Catalá, 1998).

The objective of this study was to evaluate the effect of organic fertilization in postharvest of the blueberry fruits stored in modified and conventional atmospheres.

MATERIALS AND METHODS

Raw material

Southern highbush blueberry fruits (*V. corymbosum* L. x *V. darrowii* Camp) cv. O'Neal were manually harvested on plants treated with distinct fertilization treatments from an orchard planted 2 years earlier, first harvest, located in the Nogal Experimental Station of the Universidad de Concepción, Chillán Campus (36°59' S; 72°08' W). Treated plants were randomly distributed in the orchard with four replicates per fertilization treatment (Tables 1 and 2). However, harvest was carried out for

each fertilization treatment from the total of replicates, considering that the second-year plants obtained low production (< 0.5 kg plant⁻¹), especially those organically treated.

The fruits were selected by color (> 90% blue surface), rejecting those with low caliber (< 0.5 cm), overripe and underripe, with microorganism development, soft and/or split. The selected fruits were taken to the Postharvest Laboratory in the Pilot Plant of the Departamento de Agroindustrias of the Universidad de Concepción, Chillán Campus, where the assay was prepared for storage. The total of fruits from each fertilization treatment was divided in equal parts to be stored in a modified atmosphere (MA) and a conventional atmosphere (CA). In each storage atmosphere, the fruits from the distinct fertilization treatments (total of 13, Table 1) were divided into three replicates per treatment for each day of evaluation (0, 15, and 30 d), resulting in a total of 234 plastic pots (Clamshell Driscoll's 125 g capacity) in which the blueberry fruits were deposited.

Experimental design

The behavior of fruits from plants treated with 13 different fertilization treatments and stored in two atmospheric conditions (MA and CA) in three storage periods corresponding to day 0, 15, and 30 were evaluated. A subdivided plot design with factorial arrangement 13 x 2 x 3 was applied (Table 3) for the variables percentage humidity, soluble solids, titratable acidity, pH, and percentage decay. A divided plot design with factorial arrangement 2 x 13 (Table 3) was also applied where the factors corresponded to the two atmospheric conditions (MA and CA), and fertilization type for the weight and equatorial diameter loss

Table 1. Fertilization treatments used in the blueberry orchard, first season. Chillán, Chile.

T	Fertilization ¹	Postplantation dose ¹
T1	Broiler bed 10	10 m ³ ha ⁻¹
T2	Broiler bed 20	20 m ³ ha ⁻¹
T3	Hog guano 10 (HG)	10 m ³ ha ⁻¹
T4	Hog guano 20	20 m ³ ha ⁻¹
T5	Fish filter	10 m ³ ha ⁻¹
T6	HG + Tea compost	10 m ³ ha ⁻¹ + 100 L ha ⁻¹ irrig ⁻¹
T7	HG + Bioplasm	10 m ³ ha ⁻¹ + 20 L ha ⁻¹ (6 appl.)
T8	HG + Supermagro	10 m ³ ha ⁻¹ + 100 L ha ⁻¹ irrig ⁻¹
T9	HG + Fruticrop	10 m ³ ha ⁻¹ + 5 L ha ⁻¹ (5 appl.)
T10	HG + Oiko-Bac-Nitrobio	10 m ³ ha ⁻¹ + 1 L ha ⁻¹ (2 appl.)
T11	HG + Fish oil	10 m ³ ha ⁻¹ + 1 L ha ⁻¹ irrig ⁻¹
T12	Without fertilization	-
T13	Conventional fertilization	55 kg ha ⁻¹ (urea)

T: treatment; appl.: applications; irrig: irrigation.

¹Dose frequently used by organic producers in Chile.

Table 2. Description of organic fertilizers used.

Organic fertilizer	Description
Broiler bed	Droppings composted with sawdust. Estimated nutrient input: 1.40% N, 1.85% P, and 1.55% K.
Hog guano	Composted droppings. Estimated nutrient input: 0.80% N, 0.40% P, and 0.35% K.
Fish filter	Subproduct of fish oil production, consistent with the solid surplus trapped in the filter. High biological value.
Tea compost	Extract of compost fermented in water for 7 days. Its nutrient richness depends on compost quality.
Bioplasm	Microalgae suspension (<i>F. coronella</i>). Transform minerals into organic compounds assimilable by plants that receive them in their vascular system through leaves or roots, according to whether they were applied. Contains 720 mg L ⁻¹ N, 240 mg L ⁻¹ P, and 6.22 mg L ⁻¹ K.
Supermagro	Biofertilizer standardized by CET-Yumbel, consisting in a solution of guano, water, milk, molasses, and minerals fermented for 30 to 45 days.
Fruticrop	Biostimulant base of macro and microelements, amino acids, carbohydrates, vitamins, and phytohormones. Contains 7.20% N, 3.60% P, and 4.80% K.
Oiko-Bac-Nitrobio	Biological fertilizer consisting of various strains of non symbiotic <i>Azotobacter</i> bacteria able to fix N atmospherically and make it available to be absorbed by the plant.
Fish oil	Product from pressed fish with a complex chemical composition that depends on the structure of fat acids, which vary considerably depending on the species of fish and its food.

variables. Both variables were quantified with the percent difference between day 0 and 30.

Storage atmospheres

Fruit storage in MA was carried out by packing the pots in colorless bags of selective permeability, 175 x 300 mm in size and 58 mm thick (model BB4L, Cryovac, New Jersey, USA). Before packing with a gas mixing machine (Witt model KM 100-3 MEM, Witten, Germany), a mixture of 4% O₂, 10% CO₂, with the remaining 86% N, and before emptying this gaseous mixture in the bag and sealing it, creates a vacuum in its interior to maintain the composition of the introduced gases. The bags were sealed with a sealing machine (Multivac model A 300/16, Wolfertschwenden, Germany). Fruit storage in CA contains the normal gas mixture making up the air of the Earth's atmosphere and composed of 78% N, 21% O₂, and 0.03% CO₂ (Wills *et al.*, 1998). Storage temperature for both atmospheres was 3 °C and 90% relative humidity (RH).

Variables evaluated

Percentage humidity (%HFMB). This was determined by a gravimetric method in three standard-sized fruits (14 mm equatorial diameter and 10 mm polar diameter) for each replicate of each fertilization treatment. Fruits were weighed before being put into a common drying oven (Weiss Gallenkamp 0VB-300-010N, Loughborough, UK), for 48 h at 70 °C and percentage humidity fresh matter

basis (%HFMB) of the blueberries by weight differences on an analytical scale ± 0.01 g (Denver Instrument PK-352, Denver, Colorado, USA).

Weight and equatorial diameter loss. These were determined for each fertilization treatment in three replicates, quantifying the difference with respect to weight and equatorial diameter between day 0 and 30 with an analytical balance ± 0.1 g (Denver Instrument PK-352, Arvada, Colorado, USA) and a digital calibrator (General Tools, 147, New York, USA) ± 0.01 mm. The result was expressed as a percentage of the initial value.

Soluble solids (SS), pH, and titratable acidity. These were determined in the juice of the blueberry fruits for SS with a manual thermo-compensated refractometer (Sudelab RHB-40, Santiago, Chile) on each evaluation day, while pH was determined in 2 g of macerated fruits with a pH meter (Hanna Instruments Chile HI 8521, Santiago, Chile). Then each maceration was diluted in 75 mL distilled water with an agitator (Weiser Analitica, 752A, Santiago, Chile) and pH was adjusted to 8.1-8.2 by adding NaOH 0.01 N. The result was expressed as a percentage of predominant citric acid.

Presence of pathogens. On each evaluation day, fruits with decay and presence of fungal mycelia were counted, taken to the phytopathology laboratory of the Facultad de Agronomía of the Universidad de Concepción to carry

Table 3. Results of ANOVA for experimental design of plots divided with factorial arrangement 2 x 3 x 13 and split plot experimental design with factorial arrangement 2 x 13. Period of 0-30 d.

Factors	Dependent variables						
	%HFMB	%SS	TA	pH	%Pd	%WI	%DI
A	*	*	*	ns	*	*	*
D	*	*	*	*	*	na	na
T	ns	*	*	*	*	ns	*
A x T	ns	ns	ns	ns	*	ns	ns
D x T	ns	*	*	*	*	na	na
A x D	*	*	ns	ns	*	na	na
A x D x T	ns	ns	ns	ns	*	na	na

* $P \leq 0.05$. %HFMB: percentage of humidity fresh matter basis; %SS: percentage of soluble solids; TA: titratable acidity; %Pd: percentage decay; %WI: percentage weight lost; %DI: percentage diameter lost; ns: not significant ($p > 0.05$); na: not applicable to experimental design. A: storage atmosphere; D: days of storage; T: fertilization treatment.

out pathogen culture and identification. The result was expressed as a percentage of the total fruits in each plastic container.

Statistical analysis

Angular transformation was applied to the results expressed as a percentage to normalize the data (Little and Hills, 1978). Analysis of variance was conducted with Statistica 6.0 (StatSoft, 1996) software at 95% confidence level and the Tukey test ($P \leq 0.05$) was applied when significant differences arose.

RESULTS AND DISCUSSION

Percentage humidity (%HFMB). No significant differences were found among fertilization treatments in the evaluation of this variable which averaged 81%HFMB. Furthermore, fertilization did not interact with the other factors under study (Table 3). Therefore, the fertilization source was not important in the postharvest of the stored fruits for this variable. Atmospheric factors and days in storage showed significant differences and interaction (Table 4).

Fruits stored in MA lost 4.2% humidity at the end of the assay compared to the initial value, while those stored in CA lost 8.6%. The first result coincides with what Loyola *et al.* (1996) found in highbush blueberry 'Blueray' stored for 28 d in MA at 0 °C, obtaining a 7% loss after storing fruits of highbush and rabbiteye blueberries for 2 wk at 7.2 °C.

The greater humidity content in the fruits stored in MA was due to water loss altered by the selective plastic permeability films used to store the fruits, and there was greater resistance exerted and increasing relative humidity inside the bags (Loyola *et al.*, 1996; Navarrete, 2004). Furthermore, a microatmosphere is produced inside the bags, low in O₂ and high in CO₂, reducing the respiration

rate, and thus water loss, since enzymes participating in glycolysis are inhibited (Watkins y Zhang, 1998).

Soluble solids (%SS). Significant differences were observed in the three factors under study and interaction between days of storage and fertilization treatment, as well as between storage atmosphere and days in storage (Table 3).

Fruits stored in MA did not show significant differences in SS over time, with a mean value of 14.6%. On the contrary, fruits stored in CA showed significant differences with an increase in SS from 14.6 to 16.7% (Table 4).

The increase in SS over time coincides with what Miller (1988) informed in rabbiteye blueberry, Bounous *et al.* (1997) in highbush blueberry, and Hevia *et al.* (2000) in red currant (*Ribes rubrum* L.). SSs increased over time due to fruit dehydration. Loyola *et al.* (1993) state that if greater water loss occurs then the effect is a concentration of SSs. The fruits stored in CA were statistically different than those stored in MA showing greater SS contents starting on storage day 15. MA decreases fruit dehydration by exerting greater resistance to water loss (Loyola *et al.*, 1996). This influence of MA on the lower stored fruit dehydration coincides with that observed in the SS variable previously evaluated.

There were no differences between fruits from organic fertilization treatments and fruits from conventional fertilization (T13) maintained over time (Table 5). SSs probably decreased due to the consumption of the fruit soluble sugars, but their water loss proved to have a greater sugar concentration, therefore SSs increased. However, these two effects are opposed so that SSs tend to be maintained over time (Navarrete, 2004).

Titrateable acidity. Significant differences were observed in the three factors under study and interaction between days of storage and type of fertilization (Table 3).

Table 4. Percentage humidity and soluble solids (SS) of blueberries for different types of atmosphere and days of storage.

TA	Storage period							
	Day 0		Day 15		Day 30		Mean	
	%HFMB	%SS	%HFMB	%SS	%HFMB	%SS	%HFMB	%SS
MA	83.4aA	14.7aA	82.1aAB	14.2aA	79.9aB	14.8aA	81.8	14.6
CA	83.3aA	14.7aA	82.0aA	15.5bA	76.1bB	16.7bB	80.5	15.6

TA: type of atmosphere; MA: modified atmosphere; CA: conventional atmosphere.

Capital letters horizontally different indicate differences between days of storage for the same atmosphere, and lower case letters vertically different indicate differences between types of atmospheres for the same storage day, according to Tukey test ($P \leq 0.05$).

The fruits stored in CA showed greater titratable acidity (0.41% predominant citric acid) than those stored in MA (0.37% predominant citric acid). This result is an effect of the greater dehydration of the fruits stored in CA, as was observed in the above-mentioned percentage humidity and SS variables. The same as for SS, fruit dehydration provokes an acid concentration, and consequently shows a greater value in MA.

Acidity of most fruits from organic fertilization treatments was maintained over time, the same as those with conventional fertilization (T13). Only the organic treatments T7 and T11 varied in acid content over time up to storage day 30, with an acidity increase of 0.32% to 0.56%, and 0.26% to 0.44%, respectively (Table 5).

Citric acid is the principal organic acid of the blueberry; therefore titratable acidity is attributable to a great extent

to its presence (Wiley, 1997; Hevia *et al.*, 2000; Gil, 2004). Acids are one of the energy reserves of the fruit; therefore these are used in the respiration process and converted to more simple molecules such as CO₂ and water (Wills *et al.*, 1998). As a result of respiration, acids decrease, but water loss in the fruit increases its concentration. As a consequence, titratable acidity tended to be maintained over time in most of the fertilization treatments. In other assays, an increase in acidity over time was observed, with values greater than those of the assay. Thus, in rabbiteye blueberry 'Woodard', Smittle and Miller (1988) found an increase in acidity over time of 0.52% to 0.63% in 21 days of storage at 5 °C. On the other hand, Loyola *et al.* (1996) found an increase in acidity starting on storage day 7 at 0 °C, with values varying from 0.64% to 0.85% in highbush blueberry 'Blueray'.

Table 5. Percentages of soluble solids (%SS) and citric acid (%Ac), and pH of blueberries for different fertilizer treatments and days of storage.

T	Day 0			Day 15			Day 30		
	%SS	%Ac	pH	%SS	%Ac	pH	%SS	%Ac	pH
T1	15.1abA	0.27aA	4.27abA	14.7aA	0.32aA	4.09abA	15.8abA	0.38bA	4.15aA
T2	14.7abA	0.36aA	4.13abcdA	15.7aA	0.32aA	4.22aA	17.2abA	0.42abA	4.06aA
T3	16.4abA	0.43aA	3.71dA	14.0aA	0.40aA	3.79abA	15.0abA	0.44abA	3.85abA
T4	12.4bA	0.39aA	3.69dA	14.9aA	0.40aA	3.69bA	15.0abA	0.50abA	3.51bA
T5	15.0abA	0.36aA	3.77cdA	13.8aA	0.50aA	3.64bA	15.2abA	0.49abA	3.72abA
T6	17.3aA	0.39aA	3.74cdA	15.5aA	0.38aA	3.81abA	16.2abA	0.40abA	3.79abA
T7	16.5abA	0.32aA	4.17abcA	15.1aA	0.40aAB	3.86abAB	14.9abA	0.56aB	3.68abB
T8	13.7abA	0.32aA	3.94bcdA	14.8aA	0.36aA	3.89abA	16.4abA	0.48abA	3.80abA
T9	13.3abA	0.30aA	4.25abA	14.5aA	0.38aA	3.90abA	17.6aA	0.34bA	3.92abA
T10	13.1abA	0.29aA	4.10abcdA	13.8aA	0.43aA	3.76bA	12.8bA	0.43abA	3.85abA
T11	15.5abA	0.26aA	4.40aA	14.8aA	0.38aAB	3.84abB	16.7abA	0.44abB	3.78abB
T12	13.3abA	0.34aA	4.40aA	16.2aA	0.38aA	3.89abB	16.2abA	0.37bA	3.88abB
T13	14.4abA	0.40aA	4.23abA	14.7aA	0.41aA	3.81abA	16.0abA	0.43abA	3.81abA

T: fertilization treatment; T1: broiler bed 10; T2: broiler bed 20; T3: hog guano 10 (HG); T4: hog guano 20; T5: fish filter; T6: HG + tea compost; T7: HG + Bioplasm; T8: HG + Supermagro; T9: HG + Fruticrop; T10: HG + Oiko-Bac-Nitrobio; T11: HG + fish oil; T12: without fertilization; T13: conventional fertilization (urea).

Capital letters horizontally different indicate differences between storage day for the same fertilization treatment and lower case letters vertically different indicate differences between fertilization treatments for the same storage day, according to Tukey test ($P \leq 0.05$).

Table 6. Weight and equatorial diameter loss in blueberry fruits for different storage atmospheres and fertilization treatments between 0-30 days of storage.

T	Fertilization	Weight		Diameter ¹	
		MA	CA	MA	CA
		%			
T1	Broiler bed 10	1.3aA	9.9aB	15.7ab	
T2	Broiler bed 20	3.0aA	12.0aB	14.6bc	
T3	Hog guano10 (HG)	1.9aA	12.3aB	19.8ab	
T4	Hog guano 20	3.0aA	10.9aB	22.8ab	
T5	Fish filter	1.9aA	11.2aB	7.4c	
T6	HG + Tea compost	2.5aA	9.1aB	26.5a	
T7	HG + Bioplasm	2.8aA	12.3aB	10.6bc	
T8	HG + Supermagro	2.0aA	11.1aB	23.8ab	
T9	HG + Fruticrop	2.8aA	10.4aB	26.3a	
T10	HG + Oiko-Bac-Nitrobio	1.7aA	9.8aB	9.4bc	
T11	HG + Fish oil	2.0aA	12.6aB	15.3bc	
T12	Without fertilization	1.7aA	11.7aB	16.0ab	
T13	Conventional fertilization (urea)	3.1aA	11.1aB	11.4bc	
Mean		2.3A	11.0B	15.6A	18.2B

¹Mean of the storage atmospheres.

T: fertilization treatment; MA: modified atmosphere; CA: conventional atmosphere.

Capital letters horizontally different indicate differences between storage atmospheres and lower case letters vertically different indicate significant differences between fertilization treatments, according to Tukey test ($P \leq 0.05$).

pH. Significant differences and interaction were observed between fertilization and days of storage factors (Table 3). In most fruits from organic fertilization treatments, pH was maintained over time, the same as for the fruits from conventional fertilization (T13), with the exception of T7, T11, and T12 that showed significant differences due to a decrease in pH over time (Table 5). The tendency of pH decreasing over time coincides with results found by Navarrete (2004) for rabbiteye 'Bonita' blueberry and results by Smittle and Miller (1988) for rabbiteye blueberry 'Climax' and 'Woodard'. Diffusion of CO₂ in the tissues of the fruit will decrease pH according to Navarrete (2004).

The CO₂ released in the metabolic processes of the cell reacts with water forming carbonic acid which is unstable and dissociates releasing H⁺ and HCO₃⁻ (Cabezas, 2004). The free protons would explain the pH decrease whereas HCO₃⁻ would explain titratable acidity increase.

At the start of the assay, the greatest difference in pH observed was between the fruits from the distinct fertilization treatments, being significant differences between some fruits from organic fertilization treatments with respect to those with conventional fertilization. As storage time increased there were fewer differences between fertilization treatments in relation to pH (Table 5). Therefore, the effect on pH of fertilization treatments of stored fruits decreased over storage time.

Inferior pH values have been reported in highbush blueberry by Makus and Morris (1987) with a pH of 3.14, and by Loyola *et al.* (1993) in highbush blueberry 'Bluecrop' and 'Blueray' with pH 3.45 and 3.57, respectively.

Weight loss (%). In this variable, there was no interaction between factors and significant differences were found only in the storage atmosphere factor (Table 3). Fruits from organic fertilization treatments behaved in the same way as fruits from conventional fertilization (T13) in both storage atmospheres. However, each treatment behaved in a distinct way when stored in each atmosphere (Table 6).

Fruits stored in MA demonstrated the lowest weight losses (2.3% mean), showing significant differences with the fruits stored in CA with the most weight loss (11.0% mean). The greatest weight loss in fruits stored in CA coincided with that observed by Navarrete (2004) in rabbiteye blueberry 'Bonita' and by Bounous *et al.* (1997) in highbush blueberry 'Dixi', 'Coville', and 'Darrow'.

Fruits weight loss is mainly due to water loss which is produced by a difference in vapor pressure between the fruits and surrounding air. This loss is affected by the area/volume relationship, mechanical wounds on the epidermis area, the nature of the bag area, and storage temperature (Wills *et al.*, 1998). Small-sized fruits such as blueberries show a high area/volume, therefore dehydrate more than

Table 7. Percentage of blueberry fruit decay for different fertilizer treatments, atmosphere types, and days of storage.

T	Fertilization	Modified atmosphere			Conventional atmosphere		
		Day 0	Day 15	Day 30	Day 0	Day 15	Day 30
		%					
T1	Broiler bed 10	0.0aA	9.0aB+	0.0bA+	0.0aA	0.0cA+	45.3abB+
T2	Broiler bed 20	0.0aA	2.0abA+	8.3abA	0.0aA	28.3abB+	0.0cA
T3	Hog guano10 (HG)	0.0aA	0.0bA	0.0bA+	0.0aA	0.0cA	43.0abB+
T4	Hog guano 20	0.0aA	0.0bA+	0.0bA+	0.0aA	31.3aB+	43.7abC+
T5	Fish filter	0.0aA	0.0bA	0.0bA	0.0aA	0.0cA	0.0cA
T6	HG + Tea compost	0.0aA	0.0bA	0.0bA	0.0aA	0.0cA	0.0cA
T7	HG + Bioplasm	0.0aA	0.0bA	0.0bA+	0.0aA	0.0cA	37.0bB+
T8	HG + Supermagro	0.0aA	0.0bA	0.0bA	0.0aA	0.0cA	0.0cA
T9	HG + Fruticrop	0.0aA	0.0bA+	2.7bA	0.0aA	27.7abB+	0.0cA
T10	HG + Oiko-Bac-Nitrobio	0.0aA	0.0bA+	0.0bA+	0.0aA	20.7bB+	49.0aC+
T11	HG + Fish oil	0.0aA	0.0bA	0.0bA	0.0aA	0.0cA	0.0cA
T12	Without fertilization	0.0aA	0.0bA	13.7aB+	0.0aA	0.0cA	0.0cA+
T13	Conventional fertilization (urea)	0.0aA	0.0bA	0.0bA	0.0aA	0.0cA	0.0cA

T: fertilization treatment.

Capital letters horizontally different indicate differences between days of storage for the same treatment and type of atmosphere, while lower case letters vertically different indicate differences between treatments for the same day of storage and type of atmosphere, according to Tukey test ($P \leq 0.05$).

The + sign horizontally indicates differences between types of atmospheres for the same treatment and day of storage, according to Tukey test ($P \leq 0.05$).

larger-sized fruits since they have a greater area to then have a waterproof and vapor proof waxy covering (bloom) which influences weight loss, restricting water loss by evaporation (Wills *et al.*, 1998; Cabezas, 2004; Navarrete, 2004). The fruit peduncular scar should be small and dry after being harvested given that it is a source of fruit water loss (Makus and Morris, 1987; Buzeta, 1997).

Loss of equatorial diameter (%). Significant differences were observed in the storage atmosphere and fertilization factors, but no interaction was shown between them (Table 3).

Fruits stored in CA showed a greater equatorial diameter loss (18.2% mean), differing from fruits stored in MA which showed the lowest equatorial diameter loss (15.6% mean) (Table 6). Also observed in the previous parameters was an effect of MA on stored fruits showing lower losses of initial characteristics. As it was already mentioned, MA exerts resistance to water and shows a greater CO₂ content than CA which would have inhibited dehydration and degradation of pectic substances in the stored fruits, conserving their shape, therefore, their size (Cabezas, 2004; Navarrete, 2004).

Most of the fruits from organic fertilization treatments had the same diameter as fruits from conventional fertilization (T13), except for T6 and T9 which showed the greatest diameter reductions (Table 6).

After harvesting, the size of the fruits can be altered

by water content which is maintained by osmotic forces inside the cells, by the degradation of pectic substances that weaken the cell walls, and by the cohesive forces that maintain the union between cells (Wills *et al.*, 1998). As a consequence, the fruits are not able to maintain their shape and integrity (Cabezas, 2004).

Presence of pathogens. The three factors showed differences and interactions (Table 3). The presence of pathogens was expressed as a percentage of decayed fruits. The greatest percentage of decay was in the fruits stored in CA up to storage day 30 (Table 7), while the fruits stored in MA showed low decay percentages in contrast with those stored in CA, coinciding with that observed by Cabezas (2004) in raspberry (*Rubus idaeus* L.), and by Ceponis and Cappellini (1985) in highbush blueberry. Ceponis and Cappellini (1985) point out that the bag in MA increases the CO₂ concentration in respiration, thus decreasing decay. According to Romojaro *et al.* (1996) and Wills *et al.* (1998), an atmosphere with 10% or more CO₂ slows spore germination and fungal mycelia development.

Decay in this assay was caused by *Botrytis cinerea*, *Alternaria* sp. and *Cladosporium* sp. However, the pathogen with the greatest incidence was *B. cinerea* which infected flowers, small branches, and fruits in high humidity (over 95%) conditions and with moderate temperatures between 15-20 °C (Gough, 1994). The

presence of this fungus coincides with that observed by Navarrete (2004) in rabbiteye blueberry 'Bonita', and Hevia *et al.* (2000) in red currant. The incidence of *Alternaria* sp. and *Cladosporium* sp. coincides with the results by Loyola *et al.* (1993) in highbush blueberry 'Blueray', and results by Loyola and Andrade (1993) in highbush blueberry 'Elliott'.

In general, fruits from organic fertilization treatments tended to show decay, except for T5, T6, T8, and T11 which behaved in the same way as fruits from conventional fertilization (T13), without showing decay over time and in none of the storage atmospheres (Table 7).

CONCLUSIONS

In most of the variables evaluated, there were no differences in the postharvest behavior between fruits from organic fertilization and those from conventional fertilization.

The greatest presence of pathogens occurred in the fruits from organic fertilization treatments when stored in CA, being *Botrytis cinerea* the causal agent with the greatest incidence.

Fruits stored in MA showed a better postharvest behavior than those stored in CA, showing lower weight and diameter losses, lower microorganism incidence, and a lower variation over time of soluble solids, acidity, and humidity.

RESUMEN

Comportamiento en poscosecha de frutos de arándano cv. O'Neal cultivados con distintos tratamientos de fertilización orgánica. Considerando que la demanda por productos orgánicos es cada día mayor y que Chile se encuentra alejado de los mercados de exportación de fruta fresca, frutos de arándano alto (*Vaccinium corymbosum* L. x *Vaccinium darrowii* Camp) provenientes de fertilización orgánica fueron almacenados a 3 °C y 90% HR durante 30 días, en atmósfera modificada (MA) y atmósfera convencional (CA) con el objetivo de estudiar el efecto de la fertilización orgánica en el comportamiento en poscosecha. En tres períodos se evaluaron contenido de humedad, pérdida de peso, pérdida de diámetro, sólidos solubles, acidez titulable, pH, y presencia de patógenos. El diseño experimental fue de parcelas divididas con arreglo factorial. Los resultados fueron sometidos a un análisis de varianza y se aplicó test de Tukey ($P \leq 0,05$) cuando se presentaron diferencias significativas. En la mayoría de las variables evaluadas no hubo diferencias en el comportamiento en poscosecha entre los frutos provenientes de fertilización orgánica y de fertilización convencional. Sin embargo,

la mayor presencia de patógenos ocurrió en los frutos provenientes de fertilizaciones orgánicas cuando fueron almacenados en CA, siendo *Botrytis cinerea* el agente causal de mayor incidencia. Los frutos almacenados en MA presentaron un mejor comportamiento en poscosecha que los almacenados en CA, es decir presentaron menores pérdidas de peso (2,3%), pérdida de diámetro (15,6%) e incidencia de microorganismos, y una menor variación en el tiempo de sólidos solubles (14,6 %), acidez (0,37% ácido cítrico) y humedad (81%).

Palabras clave: fertilización orgánica, atmósfera modificada, *Botrytis*, *Vaccinium corymbosum*.

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