RESEARCH



GENOTYPE × ENVIRONMENT INTERACTION IN CANOLA (Brassica napus L.) SEED YIELD IN CHILE

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ABSTRACT

Genotype x environment (G × E) interaction in canola (*Brassica napus* L.) cultivar seed yield is unknown in Chile. The interaction was performed with the SREG (Sites Regression) model. Two experiments were conducted in five and thirteen environments in the 2008-2009 season in Central South Chile. The experimental design was a randomized complete block (RCBD) in each environment with four replicates and 26 open-pollinated or hybrid canola genotypes in Experiment 1, and RCBD with three replicates and 17 genotypes in Experiment 2. ANOVA was used to determine the significance of the G × E interaction. Biplots were used to graphically interpret and determine the best cultivar in each environment and the corresponding mega-environments. The G × E interaction was significant for seed yield in many locations in one cropping season. Most of the analyzed seed yield variation was due to environment and $G \times E$ effects. Principal components (PC1 and PC2) of the Sites Regression (SREG) model, with five and eight environments, accumulated 74.5% and 61.1% of the total variation, respectively. Two mega-environments. Six of the evaluated cultivars, all hybrids except 'Goya', were superior. The mean vs. stability analysis indicated that the Monalisa hybrid had the highest yield and was the most stable cultivar across all environments. Although the information is for only 1 yr, results could change with data from several years of experimentation. Hence, the study was carried out in many locations in order to provide validity to the results.

Key words: SREG, GGE, MET, interaction, biomass, Brassica napus.

Worldwide canola production is approximately 50 million tons and covers a total area of 27 million hectares. The main producers are China, 49%; the Economic European Community, 25%; Canada, 12%; and India, 9% (FAOSTAT, 2010). The total area of canola in Chile in the 2008-2009 season was 25 135 ha which produced oil and meal for the bovine and fish food industries (ODEPA, 2009).

The environment is the sum of all external conditions affecting cultivar growth and development. Soil texture, pH, depth, organic matter, fertility, diseases, and insects contribute additional variability to the environment (Roozeboom *et al.*, 2008). The genotype \times environment interaction (G \times E) is the response of each cultivar to

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variations in the environment (Crossa et al., 1991). The $G \times E$ interaction has been one of the principal subjects of study in breeding, allowing the generation of different methodologies for genetic improvement. It has also been a constant worry for breeders, especially when the magnitude of $G \times E$ is large, since this impedes the selection and recommendation of stable cultivars, as well as slowing selection advancement (Rodríguez et al., 2002). In many countries, before a cultivar is released for production, government authorities usually require at least 3 yr of performance tests in different environments. There are two possibilities to develop cultivars with low G × E interaction: subdivide areas in relatively homogeneous regions where cultivars need specific adjustment, or generate highstability materials adapted to a wide range of environments. The ideal cultivar would be the one with high seed yield and high stability when evaluated across different environments (Yan et al., 2007). Multi-environment trials must be established mainly to identify the best cultivars for a location and then determine if locations can be established as mega-environments (Yan et al., 2000).

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Zobel *et al.* (1988) compared traditional statistical analyses such as ANOVA, Principal Components Analysis (PCA), and linear regression to demonstrate that they were not always effective in analyzing test information in multiple environments. ANOVA is an additive model that describes principal effects and determines if the G \times E interaction is significant, but it does not discuss in depth the reasons for this. Principal Components Analysis is a multiplicative model that does not contain the origin of the variation for the principal additive effect of the cultivar or the environment, and does not precisely analyze the interaction. The linear regression method uses the environment means for which estimates are frequently weak or poor and gives information about a small fraction of the total variation generated by the G \times E interaction.

The biplot is a useful tool to visually evaluate and interpret cultivar response patterns, environments, and the $G \times E$ interaction. The biplot is a graphical representation of the simultaneous behavior of two variables. Biplots were originally proposed by Gabriel (1971) and are useful to analyze and summarize a great quantity of information through graphing (Crossa et al., 1991; Gauch, 2006). The biplot is also used to interpret analysis results of the Sites Regression (SREG) model of the information obtained in the multi-environment trials (MET). The genotype and $G \times E$ interaction (GG \times E) are important factors in selecting cultivars, and they constitute the sources of variation in the SREG model to analyze MET information. These factors are graphically estimated by the GGE biplot in which both cultivar and environment are graphically visualized (Yan et al., 2000; Yan, 2001). This model was proposed to explore cultivar response to specific environments in which the main effects of cultivars form a part of the residual (Yan et al., 2000). An exclusive merit of this model is that it allows grouping environments with similar behavior and graphically identifies which cultivar has the highest potential in each environment subgroup.

The study of GG \times E for canola (*Brassica napus* L.) is important for Chilean agriculture due to the fact that all commercialized cultivars in the country originate from other countries where environmental conditions are different. On the other hand, there is no network of coordinated tests through an official organization which can objectively determine the adaptation of canola cultivars to different locations and optimize the use of cultivars available in Chile.

The objective of this study was to evaluate $G \times E$ interaction for winter canola cultivars. Specific objectives included the characterization of 26 winter canola cultivars in different dryland locations in the Bío Bío, La Araucanía, and Los Lagos Regions in Chile during the 2008-2009 season, demonstrate the usefulness of the SREG and $GG \times E$ models, and compare the performance potential between hybrid and open-pollinated cultivars.

MATERIALS AND METHODS

Experiments were conducted in three regions of Central South Chile where canola is currently grown. Two experiments were conducted during the 2008-2009 season. Experiment 1 had 26 canola genotypes made up of hybrid cultivars: Monalisa, Hornet, Rohan, Exagone, Brutus, Tadeus, SW5, Dimensión, Hammer, Tassilo, Tuarus, Triangle, Lilian, Artus, and Cuillin; open-pollinated cultivars: Goya, Livius, Cult, Favorite, Liprima, Sunday, Compact, Galileo, Tatra, Vision, and Coronet. All of these were evaluated in five environments (Chillán: 36°35' S, 72°04' W; El Carmen: 36°56' S, 72°00' W; Los Ángeles: 37°27' S, 72°18' W; Gorbea-A: 39°05' S, 72°40' W; and Osorno: 40°24' S, 73°10' W). Experiment 2 had 17 genotypes made up of hybrid cultivars: Monalisa, Hornet, Exagone, SW5, Dimensión, Hammer, Lilian; and open-pollinated cultivars: Goya, Livius, Cult, Favorite, Liprima, Sunday, Compact, Galileo, Tatra, and Vision. These were evaluated in thirteen environments (Cañete: 37°52' S, 73°24' W; Collipulli: 37°59' S, 72°14' W; Gorbea-B: 39°05' S, 72°38' W; Lautaro: 38°30' S, 72°30' W; Máfil: 39°37' S, 73°03' W; Mulchén: 37°43' S, 72°84' W; Paillaco: 40°09' S, 72°42' W; Victoria: 38°15' S, 72°17' W; Chillán: 36°35' S, 72°04' W; El Carmen: 36°56' S, 72°00' W; Los Ángeles: 37°27' S, 72°18' W; Gorbea-A: 39°05' S, 72°40' W; and Osorno: 40°24' S, 73°10' W). For each cultivar, the seeding rate was calculated with 1000-seed weight as well as germination percentage to achieve a plant density of 25 to 30 plants m⁻² for hybrids and 30 to 40 plants m⁻² for open-pollinated cultivars.

Experimental design and agronomic management of experiments

The experimental design for both Experiments 1 and 2 was a randomized complete block design with four and three replicates, respectively. In Experiment 1, every experimental unit consisted of six 5-m rows with 0.3 m spacing. The four center rows were harvested and 0.5 m at the end of every row was discarded. The experimental unit used in Experiment 2 consisted of five 5-m rows of 5 m with 0.35 m spacing.

Traditional tillage was employed at the Chillán, Los Ángeles, and Gorbea locations, and no-tillage before planting in El Carmen and Osorno. Seeding depth was 4 cm in every location. Fertilizers and application rates were adjusted according to soil tests. Weeds, insects, and fungi were controlled by applying the following products: Herbicides: metazachlor 2.3 L ha⁻¹ [(N-(2,6dimethylphenyl)-N-(1pyrazolylmethyl) chloroacetamid)], 0.5 L ha⁻¹; fungicides: metconazole (1RS,5RS;1RS,5SR)-5-(4-chlorobenzyl)-2,2-dimethyl-1-(1H-1,2,4-triazol-1-ylmethyl)ciclopentanol) triazole; and insecticides: lambda-cyhalothrin 0.35 L ha⁻¹ [(carboxylate of (R+S)– α cyano-3-phenoxybenzyl-(1S+1R)-cis-3-(Z-2-chloro-3,3,3trifluoroprop-1-enyl)-2,2dimethylcyclopropane] (AFIPA, 2009-2010).

Seed yield was calculated in the four center rows of each experimental unit and 0.5 m of plants were discarded from the end of the rows. Biomass samples were taken from a 0.6 m² area within each plot where plants were cut at the stem base.

Statistical analysis

Information obtained in each environment was analyzed separately and combined with the General Linear Model (GLM) procedure of the SAS statistical software (SAS Institute, 2007). The combined analysis was performed with "environment" as a random effect and "genotype" as a fixed effect. Mean separation was calculated according to Tukey's test ($P \le 0.05$). A linear regression was also carried out to determine if biomass was related to seed yield.

Analysis of the cultivar × environment interaction

The matrix centered on the columns generated by the environment with the $G \times GE$ information was subjected to singular value decomposition (SVD) where every element in the matrix was estimated with the following equation:

$$E(Y_{ij}) = \mu + \beta_j + \sum_{k=1}^k \lambda_k \gamma_{ik} \delta_{jk}$$

where $E(Y_{ij})$ is the estimated value of genotype *i* in environment *j*; μ is the general mean; β_j represents the principal effect of the environment; k is the principal component (PC) number needed to describe G × GE; λ_k is a constant of variation or singular value for the *k*th PC (PC_k); γ_{ik} and δ_{jk} , are the values of the *i*th genotype and *j*th environment, respectively, for PC_k. Singular value decomposition was achieved by a factor on a large scale *f* to obtain alternative values of genotype ($n_{ik} = \lambda_k \lambda_{ik}$) and environment ($m_{ik} = \lambda_k \delta_{jk}$). Singular value decomposition from the mean GG × E forms a matrix containing *n* genotype points and *m* environment points. The simplest symmetrical scale one (f = 0.5) was chosen for the analysis (Yan and Rajcan, 2002). The statistical theory of this model has been described in detail by Yan *et al.* (2007).

With this model and matrix algebra, results of the interaction were obtained. The matrix was constructed from seed yields obtained for each replicate, genotype, and environment. Principal component analysis (PCA) was conducted to group and determine linear relationships among them. A vector was then assigned to each eigenvalue from each PC. Next, with all the opposing vectors corresponding to genotypes and environments, an orthonormal base (space of points) was constructed to predict a genotype seed yield in a certain environment, which was then projected graphically on the plane. The association among environments and genotypes in these analyses allowed determining the adaptation, nature, and/or magnitude of the G x E interaction given by the dependence and linear association among them, and grouping them in the same PC.

The SREG model biplot (GGE biplot) were interpreted according to Kempton (1984), Zobel *et al.* (1988), Crossa (1990), Crossa *et al.* (1991), Vargas and Crossa (2000), Franco *et al.* (2003), Yan *et al.* (2000; 2007), Sabaghnia *et al.* (2008), and Yang *et al.* (2009). All the biplots shown in this study were obtained with the GGE biplot software (Yan, 2007).

RESULTS AND DISCUSSION

Seed yield in Experiment 1 showed significant differences for both the genotypes and $G \times E$ interaction ($P \le 0.05$) (Table 1). The highest seed yield was obtained by the Monalisa, Hornet, Rohan, Exagone, Brutus, Tadeus, SW5, and Dimensión hybrids, along with the open-pollinated cultivar Goya which had a mean yield similar to the hybrids (Table 2). Other authors have reported similar results when comparing hybrid and open-pollinated cultivars (Ortegón *et al.*, 2006; 2007). Seed yield is a complex character that includes several components such

Table 1. Mean squares for seed and biomass yield of 26 canola genotypes cultivated in five environments in Chile in 2008-2009.

SOV	df	Seed yield	df	Biomass yield	df
G	25	1 111 614*	25	48 253 720	25
G×E	100	429 394*	100	35 578 175	100
Error	370	234 634	371	35 780 621	121
CV%		18		50	

*Significant when $P \le 0.05$, CV: coefficient of variability, SOV: sources of variation.

	Environment											
Genotype	Chillán	El Carmen	Los Ángeles	Los Ángeles Gorbea		Mean						
	kg ha ⁻¹											
Monalisa	3550	3724	2634	2547	3857	3262						
Hornet	3210	3398	2518	2054	3940	3024						
Rohan	2977	3459	2761	1580	3938	2943						
Goya	3516	3491	2638	1493	3400	2908						
Exagone	3025	3156	2332	1706	3767	2797						
Brutus	3342	2812	2509	1616	3685	2793						
Tadeus	2836	2690	2668	1780	3725	2740						
SW5	2819	2769	2625	1609	3686	2702						
Dimensión	3663	3056	2729	1094	2956	2700						
Livius	3359	2702	2678	1396	3038	2635						
Cult	3169	2784	2530	1555	3085	2624						
Hammer	3722	2879	2338	956	3050	2589						
Tassilo	2703	2383	2537	1684	3613	2584						
Taurus	3025	3092	2214	1524	2949	2561						
Favorite	3149	2989	2319	1339	3000	2559						
Triangle	1799	2980	2639	1997	3327	2548						
Lilian	3418	2315	2199	1302	3420	2531						
Liprima	3216	2597	2348	1587	2777	2505						
Sunday	3308	2272	2119	1474	3318	2498						
Compact	2823	2623	2601	1282	3154	2497						
Galileo	2901	3027	2392	1371	2757	2490						
Artus	2333	3165	2202	1474	3090	2453						
Cuillin	2491	2129	2469	1395	3369	2371						
Tatra	2913	2652	2455	1173	2511	2341						
Vision	2523	2611	2041	1216	3099	2298						
Coronet	2794	1566	2328	1292	2944	2185						
Tukey's (0.05) =	576.6											

Table 2. Seed yield of 26 canola genotypes in five environments cultivated in Chile in 2008-2009.

as plant density, number of siliques per plant, number of seeds per silique, and seed weight. For this reason, seed yield is very variable and depends on the cultivar, the environment where it is grown, and the environment selected for the cultivar (Diepenbrock, 2000; Nassimi *et al.*, 2006).

In Gorbea, it was observed that in both hybrids and open-pollinated cultivar plant density was strongly affected by frost heaving during seedling emergence and establishment. Canola needs rapid germination, emergence, and establishment to achieve uniform plant density and develop roots that are sufficiently deep before the start of autumn frosts when the risk of frost heaving increases, generally after 15 May (Diepenbrock, 2000). Frost can cause damage in the cotyledon and seedling stage, but the plant is capable of tolerating very low temperatures once it reaches the rosette stage (Thomas, 2003). Winter canola can be exposed to temperatures of up to -20 °C for short periods (FAOSTAT, 2010). Minimum temperatures registered during the emergenceestablishment period fluctuated between -4.2 and -7.2 °C in Gorbea. On the other hand, the experiment was established in an area that had been left untilled for several years, and tilling did not properly incorporate the excess residue into the soil, thus interfering with seedling establishment and contributing to plant loss during this period. In Gorbea, low density was probably one of the factors that determined the lowest seed yield with a mean of only 1520 kg ha⁻¹ for this environment as compared to Osorno with the highest mean seed yield of 3287 kg ha⁻¹. In addition, high seed shattering was observed in Gorbea (approximately 20 to 30% for all cultivars). Drought at the end of the maturity phase along with high temperatures increased seed shattering.

Nielsen (1996) reported seed yields of 538 and 3416 kg ha-1 in winter canola in Northeastern Colorado (USA) with 249 mm and 521 mm of water, respectively. This would explain higher seed yields in Osorno where soil moisture was adequate during seed development and maximum air temperatures were lower than in Los Ángeles and Chillán. Temperatures above 25 °C in Chillán were recorded in full bloom in September causing seeds to mature faster and resulting in lower seed weight. In Los Ángeles and Chillán, canola reached maturity at the beginning of December and was harvested first. ANOVA did not show any significant differences for biomass yield for both the main effect of genotype and $G \times E$ interaction (P >0.05) (Table 1). This is probably because ANOVA could not detect differences because of the high coefficient of variation (50%). Cultivars that exhibited high seed yield, such as 'Monalisa' and 'Goya', also had high biomass yield although significant differences among cultivars were not detected. On the contrary, cultivars with low biomass exhibited lower seed yield (Table 3). Produced biomass or dry matter supports the weight of plant siliques, thus increasing growth to mobilize stored photosynthates (Major et al., 1978). When hybrids and open-pollinated cultivars were compared, hybrids had higher biomass yield. Of all the environments under study, Gorbea exhibited the lowest biomass yield, and cultivar production was lower in this environment, which indicates a high correlation between biomass yield and seed yield. Differences observed among hybrid cultivar as compared to open-pollinated cultivar seed yields would be associated with the ability of hybrids to accumulate more biomass per surface unit and produce a higher number of siliques per plant (Radoev et al., 2008). In Osorno, canola plants were taller than in the other four environments under study, mainly due to a greater availability of soil water and more suitable temperatures. Optimum temperatures for canola development range between a minimum of 15 to a maximum of 25 °C although the base temperature for development is 5 °C (FAOSTAT, 2010). Seed yield and biomass are random variables in the regression model with an association of r = 0.44.

Evaluation interaction $G \times E$ with SREG model and GGE biplot

Results are discussed separately for both experiments; first, the cultivar with the best yield in each environment is indicated followed by cultivar mean performance and stability.

Experiment 1: 26 cultivars in five environments. Combined ANOVA detected significant differences in environments (E), genotypes (G) and $G \times E$ interaction (Table 4). Of the total sum of squares for $E + G + G \times E$,

Га	ble 3. Mean biomass yield of 26 canola genotypes in
	five environments (Chillán, El Carmen, Los Ángeles,
	Gorbea-A, and Osorno) cultivated in Chile in 2008-
	2009.

Genotype	Biomass yield
	kg ha-1
Artus	10 988
Brutus	12 059
Compact	10 749
Coronet	8 915
Cuillin	12 289
Cult	10 676
Dimensión	12 802
Exagone	13 522
Favorite	17 032
Galileo	11 692
Goya	12 374
Hammer	11 688
Hornet	13 407
Lilian	10 674
Liprima	9 800
Livius	10 351
Monalisa	13 138
Rohan	11 982
SW5	12 366
Sunday	12 327
Tadeus	11 355
Tassilo	11 602
Tatra	10 297
Taurus	11 918
Triangle	11 968
Vision	10 275
Tukey's (0.05) =	NS

65.8% was the result of the environment effect, so that this factor must be further analyzed. The magnitude of variation attributed to $G \times E$ interaction was more than the one for genotype, suggesting the possibility of grouping environments according to $G \times E$ interaction (Yan *et al.*, 2000; Ma *et al.*, 2004).

The first two PCs explained 74.5% (PC1 = 45.8%, PC2 = 28.7%) of the total GGE variation with the centered model = 2 and SVP = 1. The proportion of the variation explained in this study (74.5%) was considered acceptable. Although it is not clear how much of the variation must be represented by the first two principal components involved in the biplot, recent publications indicate that at least both PCs must accumulate more than 60% of the variation generated by (G × GE) to interpret mega-environments

SOV	df	SS	Variation $(\%)^1$	Mean squares	F Value	P > F
E	4	194 969 238	65.84	48 742 309	207.74	< 0.0001
Rep/E	15	30 417 091	10.27	2 027 806	8.64	< 0.0001
G	25	27 790 355	9.38	1 111 614	2.59	< 0.0001
G×E	100	42 939 453	14.50	429 394	1.83	< 0.0005
Error	370	296 116 138		234 634		

Table 4. Sum of squares (SS) and mean squares of environments (E), genotypes (G), and sources of variation (SOV) for combined ANOVA for seed yield of 26 canola genotypes cultivated in five environments.

¹Percentage of sum of squares of $E + \text{Rep}/E + G + G \times E$.; Rep: replicate.

with some validity (Yang et al., 2009). One explanation of this result was that all genotypes used are winter canola types with closely related pedigrees, and the genetic base for winter canola genotypes is also not very extensive, which may influence the $G \times E$ interaction (Long *et al.*, 2007). The cvs. Rohan, Hornet, and Exagone were better adapted to the coldest and more humid southern environments, that is 'Gorbea' and 'Osorno', than to the hotter and drier northern location of Chillán where 'Dimensión' and 'Hammer' performed better. However, 'Monalisa' had the best seed yield in four of the five studied environments, and demonstrated the good seed yield stability of this genotype. To determine 'which-won-where', or which genotype is the best in which environment, an SREG analysis was conducted with a biplot as a visual tool. The GGE biplot is an effective visual tool to analyze mega-environments (Yan et al., 2000) consisting of an irregular polygon and a set of lines drawn from the biplot origin intersecting each of the sides at a right angle (Yan et al., 2007). The genotypes that are the farthest away from the biplot origin are located in the vertices of the polygon in different directions in such a way that all the studied genotypes are contained inside the polygon. The line running from the biplot origin intersects perpendicularly with the side of the polygon representing a hypothetical set of environments in which genotypes perform equally (Yan and Rajcan, 2002). If the studied environments are located in different sectors, this means that different genotypes obtained the highest seed yield in different environments (Yan *et al.*, 2007). The biplot in Figure 1 is based on seed yields shown in Table 2. The five environments were located in two areas of the biplot marked by two different cultivars, and 'Monalisa' had the highest seed yield in all environments except Chillán. The lines perpendicular to the sides of the polygon divide the biplot into five sectors, but the five evaluated environments were located in only two of the areas or mega-environments.

An interesting characteristic of the GGE biplot visualization is that cultivars in the vertices of the polygon are extreme cultivars with either the highest or lowest seed yield for each mega-environment made up of the original environments located in that area of the biplot (Yan *et al.*, 2007). The Gorbea, Osorno, El Carmen, and Los Ángeles environments were grouped in one



Figure 1. GGE biplot for 26 canola genotypes cultivated in five environments: Chillán, El Carmen, Los Ángeles, Gorbea-A, and Osorno in 2008-2009 indicating "which-won-where" pattern for seed yield. PC1 and PC2: Principal Components 1 and 2, respectively.

mega-environment with 'Monalisa' and 'Hammer' in the vertices of the polygon suggesting that these cultivars had the highest performance in these environments. The Chillán environment was located in a different area of the biplot with 'Hammer' and 'Dimensión' suggesting that they had a high performance.

Interpretation of the biplot suggests that there are three possible outstanding cultivars, Monalisa, Goya, and Hornet for the mega-environment represented by Gorbea, Osorno, El Carmen, and Los Ángeles, as well as two cultivars in Chillán, 'Dimensión' and 'Hammer'. Information generated in multiple years and environments is essential to decide if an environment or a region can be objectively divided into different mega-environments (Yan and Rajcan, 2002). Repeated locations are necessary, but not sufficient, to establish different mega-environments since cultivar response can change from year to year (Yan and Rajcan, 2002; Yan et al., 2007; 2009). Some authors have reported that the 'which-won-where' pattern is not repeatable in many cases, so that it is not possible to separate cultivation areas in real mega-environments (Navabi et al., 2006).

To determine average potential performance and genotype stability, these factors were analyzed with the "Average Environment Coordination" method (AEC) (Yan, 2001; Yan and Rajcan, 2002; Yan *et al.*, 2007). In this method, the average performance of environments is defined as the mean of PC1 and PC2 values of all environments represented by the small circle in Figure 2, and a single arrow line is drawn through the environment average and biplot origin. This AEC abscissa is the "average environment axis". The AEC coordinate axis, AEC Y-axis, is the line that passes through the origin and is perpendicular

to the AEC abscissa. Unlike the X-axis, which has a single direction towards the best cultivar, the Y-axis is indicated by the double arrow line. Genotypes located away from the AEC abscissa are less stable than those near the top or close to the AEC line. The Y-axis separates genotypes with lower and higher than average performance from the general genotype mean. Genotypes with a mean seed yield higher than the general mean for all genotypes were 'SW5', 'Tadeus', 'Dimensión', 'Brutus', 'Exagone', 'Goya', 'Rohan', 'Hornet', and 'Monalisa' whereas genotypes with a mean seed yield lower than the general mean were 'Hammer', 'Livius', 'Favorite', 'Taurus', 'Liprima', 'Lilian', 'Sunday', 'Compact', 'Tassilo', 'Artus', 'Triangle', 'Vision', 'Tatra', 'Cuillin', and 'Coronet' (Figure 2). It is important to emphasize that this analysis method does not statistically differentiate genotypes and since hypothesis tests are not used in the analysis, conclusions are drawn by observing the biplot (Yang et al., 2009).

On the other hand, genotype stability is important since it indicates if a high-yielding genotype in one environment maintains its relative ranking across environments. A very long line projected from the AEC coordinate, independent of its direction, represents a high G x E interaction. The mean of these genotypes can be more changeable in different environments and can then be considered as having lower stability. 'Monalisa' was the genotype with the highest stability and best performance (Figure 2).

Genotypes in this study were classified according to the methodology suggested by Yan *et al.* (2007). Genotypes in the first group had the highest yield potential and high stability, and were therefore described as superior genotypes: 'Monalisa', 'Hornet', 'Rohan', 'Exagone', 'Brutus', 'Goya', and 'SW5'. It must be emphasized that





all cultivars in this group were hybrids except Goya. The second group included 'Dimensión' and 'Tadeus' with high yield potential, but low stability, that is, high seed yield only in some specific environments. The third group included cultivars with low seed yield potential, but high stability, a comparatively lower yield in all the evaluated environments and would therefore, not be recommended for any of the studied environments: 'Coronet', 'Vision', 'Tatra', 'Compact', 'Galileo', 'Sunday', 'Liprima', 'Lilian', 'Taurus', 'Favorite', and 'Cult'. 'Coronet' was classified as low-yielding and highly stable. It is an old cultivar, early flowering and maturing, of lower height, biomass, and seed yield. All the cultivars classified in this group were open-pollinated except for 'Lilian' and 'Taurus', the latter being an old hybrid that will probably be promptly withdrawn from the market because of its susceptibility to Alternaria spp. The fourth group cultivars had low seed yield only in some of the environments: Tassilo, Triangle, Hammer, Artus, Cuillin, and Livius.

Experiment 2: 17 cultivars in 13 environments. Combined ANOVA detected significant differences for environments (E), genotypes (G), and $G \times E$ interaction (Table 5). Of the

total sum of squares of $E + G + GG \times E$, 82.1% was the result of the environment, so that it is necessary to analyze how this factor influences the evaluated cultivars. The magnitude of the variation attributed to $G \times E$ also suggests the possibility of grouping environments in terms of the G × E interaction (Yan *et al.*, 2000). These results coincided with those obtained in Experiment 1 (Table 6).

The first two PCs explained 61.1% (PC1 = 46.7%, PC2 = 14.4%) of the total GGE variation with the centered model = 2 and SVP = 1. Analyzing these cultivars with only 61% of the total variation implies the risk of ignoring important aspects of the variation caused by $G \times E$ interaction. If we compare these results with those of the previous analysis with 26 cultivars and five environments, the ranking and grouping of cultivars in mega-environments, and the selection of high performance cultivars, they were similar. This is the main reason for discarding the G × E variation of 39% and not using the third PC in a 3-D biplot (Yang et al., 2009). In addition, other researchers, such as Kaya et al. (2006) and Roozeboom et al. (2008) also used only two PCs with a cumulative variation of 62% for the construction of the GGE biplot in 25 cultivars of Triticum aestivum L.

Table 5. Sum of squares (SS) and mean squares of environments (E), genotypes (G), and sources of variation (SOV) for combined ANOVA for seed yield of 17 canola genotypes cultivated in 13 environments.

SOV	df	SS	Variation $(\%)^1$	Mean squares	F Value	P > F
E	12	744 166 880	82.15	62 013 906	271.9	< 0.0001
Rep/E	26	23 452 404	2.58	902 015	3.9	< 0.0001
G	16	57 580 725	6.35	3 598 795	8.4	< 0.0001
G×E	188	80 573 704	8.89	428 583	1.9	< 0.0001
Error	408	905 773 714				

¹Percentage related to sum of squares of E + Rep E + G + G × E.; Rep: replicate.



Figure 3. GGE biplot for 17 canola genotypes cultivated in 13 environments: Chillán, El Carmen, Los Ángeles, Gorbea A, Gorbea B, Mulchén, Cañete, Collipulli, Victoria, Lautaro, Máfil, and Paillaco in 2008-2009 indicating "which-won-where" pattern for seed yield. PC1 and PC2: Principal Components 1 and 2, respectively.

Table 6. Seed yield for 17 canola genotypes in 13 environments cultivated in Chile in 2008-2009 season, Experiment 2.

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Genotypes	Caffet	e collip	chill?	IL BICS	str Gorb	eis Goth	eice 105	Int Lant	art Mali	Mult	ne Osor	no Paille	victo	Mean
							— kg	ha ⁻¹ —						
Monalisa	6569	3761	3713	3489	2563	2640	2741	3539	5449	4550	4236	5171	4120	4041
Hornet	5975	3781	3293	3420	2166	2647	2429	4422	5253	2770	3866	5130	3393	3734
Exagone	6040	3999	2984	3393	1802	2858	2362	3615	5064	2240	3967	5038	3738	3623
Goya	5272	4069	3804	3791	1603	1936	2536	2129	4390	3195	3536	5695	3062	3463
SW5	5348	3123	2746	2714	1740	2208	2520	3002	4579	4063	3776	5883	3059	3443
Dimensión	5880	3996	3654	3330	1166	1662	2510	2751	4269	2311	3176	5434	2771	3301
Liprima	5384	4104	3163	2541	1664	2059	2592	1976	4522	2454	3092	5317	3945	3293
Cult	5071	3393	3164	2901	1564	1716	2427	2809	4342	3328	3316	4807	3608	3265
Livius	5019	3712	3492	2977	1597	1734	2411	2757	4536	2271	2982	4880	2858	3171
Lilian	4637	3915	3438	2512	1439	1507	2199	2256	4673	-	3624	4733	3073	3167
Vision	5502	3946	2294	2906	1268	1509	2125	2754	4630	2724	2904	4989	3464	3155
Galileo	4934	3671	2757	3381	1322	2046	2233	2965	3889	2414	2878	4979	3325	3138
Compact	5251	3122	2890	2657	1538	1610	2370	2502	4180	-	3214	5044	3269	3137
Favorite	5104	3611	2930	2857	1602	1985	2401	2997	4098	-	3123	3465	3009	3098
Hammer	5346	2982	3631	3059	1081	1769	2402	2745	4213	2127	3198	4737	2247	3041
Sunday	5256	3070	3357	2335	1579	1540	2014	2460	3756	1702	3381	4665	3071	2937
Tatra	4487	3140	2848	2601	1218	1772	2289	2647	3544	-	2526	4526	2966	2880
Tukey's	(0.05)=	466												



Figure 4. "Mean vs. Stability" biplot for 17 canola genotypes cultivated in 13 environments: Chillán, El Carmen, Los Ángeles, Gorbea-A, Gorbea-B, Osorno, Mulchén, Cañete, Collipulli, Victoria, Lautaro, Máfil, and Paillaco in 2008-2009. PC1 and PC2: Principal Components 1 and 2, respectively. AEC: Average Environment Coordination.

In order to visualize the same cultivar performance as in Experiment 1 in a greater number of environments, cultivar trial results from Agrosearch Ltda. were used in addition to the five studied environments. Experiments 1 and 2 were not combined into a single analysis because the variances were not homogeneous, and harvest and seed cleaning methods were different. The 17 cultivars evaluated in 13 environments were grouped in two sectors of the biplot with different superior cultivars on the vertices of the polygon (Figure 3). 'Monalisa' was the cultivar with the highest yield in 12 of the 13 environments and 'Exagone' and 'Hornet' had the highest yield only in the Lautaro environment. These results coincide with those from Experiment 1 in which 'Monalisa' had the highest yield in most environments. Figure 4 shows that the highest seed yield means were for 'Goya', 'SW5', 'Exagone', 'Hornet', and 'Monalisa', and the lowest means for 'Cult', 'Liprima', 'Dimensión', 'Lilian', 'Compact', 'Favorite', 'Hornet', 'Sunday', and 'Tatra'. 'Monalisa' was the most stable and showed the highest performance.

According to the results of this study during the 2008-2009 season, the 'Monalisa' hybrid showed the best seed yield potential in all environments with a high overall stability, which would classify it as a stable high performance cultivar. The cultivars with high seed yield in only some environments and low stability were SW5, Goya, Exagone, and Hornet. The cultivars with low seed yield potential in all environments were Vision, Dimensión, Liprima, Tatra, Livius, and Galileo. The cultivars in the fourth group that had a low seed yield in only some environments were Lilian, Favorite, Cult, Sunday, Vision, and Hammer. It is widely accepted that quantitative characters such as seed yield are controlled by multiple genes and their expression depends on the environment (Long *et al.*, 2007).

CONCLUSIONS

It was concluded that seed yield in canola grown in Chile is strongly influenced by the genotype \times environment interaction of the total variation. The environment and genotype \times environment interaction were the most important with 65.8 and 14.5% of the variation, respectively, when evaluated in five environments, and 82.2 and 8.9% of the variation, respectively, in 13 environments.

Of the 26 evaluated genotypes, six cultivars were welladapted to the environments of El Carmen, Los Ángeles, Gorbea A, and Osorno, as well as two which were better adapted to Chillán in the 2008-2009 season.

In this study, 'Monalisa' had the highest seed yield and stability in most of the analyzed environments according to SREG analysis and GGE biplot. 'Hornet', 'Goya', and 'Exagone' also had an outstanding performance.

According to the results, the cultivars most adapted to the canola production region of Chile would be the 'Monalisa', 'Hornet', and 'Exagone' hybrids, and the open-pollinated Goya cultivar.

The conclusion is based on only 1 yr of experimentation, so that results could change if more data were included covering a longer period of time.

ACKNOWLEDGEMENTS

Funding for this research was provided by FIA (Fundación para la Innovación Agraria), project FIA-PI-C-2007-1-A-008, Chile. Authors acknowledge the valuable collaboration of ALISUR S.A., Ricardo Montesinos Iroume, Molinera Gorbea Ltda., Biosemillas Ltda., Hernán Martínez Chavarría, and Waldo Cerón of Agrosearch Ltda. for providing 13 more study sites. We also thank the technicians and students for plot planting, management, data collection, and analysis, especially Wilson González Saavedra and Alejandro Solis Fuentes.

RESUMEN

Interacción genotipo × ambiente en el rendimiento de semilla de raps (Brassica napus L.) en Chile. La interacción genotipo x ambiente en raps (Brassica napus L.) en Chile es desconocida. Para su determinación se utilizó el modelo SREG (regresión de sitios). Dos experimentos fueron establecidos en la temporada 2008-2009. El diseño experimental en cada ambiente fue de bloques completos al azar con cuatro repeticiones y 26 cultivares de polinización abierta o híbridos de raps para el Experimento 1 y de bloques completos al azar con tres repeticiones y 17 cultivares para el Experimento 2. Se utilizó un ANDEVA para determinar la significancia de G × E. Se usó el biplot para la interpretación gráfica y determinación del cultivar superior en cada ambiente y determinar el correspondiente mega ambiente. En una temporada y para las localidades evaluadas la interacción G x E fue significativa para rendimiento de semilla. La variación en el rendimiento de semillas determinada por el análisis fue debido al efecto del ambiente y de la interacción G x E. Los componentes principales (PC1 y PC2) del modelo SREG con cinco y trece ambientes acumularon 74,5% y 61,1% del total de la variación, respectivamente. Se formaron dos mega ambientes, un ambiente representado por Chillán y el otro formado por el resto de los ambientes estudiados. Seis de los cultivares evaluados fueron superiores y todos fueron híbridos excepto 'Goya'. El análisis del promedio vs. estabilidad indicó que el híbrido Monalisa fue el de mayor rendimiento y el más estable evaluado a través de todos los ambientes. La información presentada sólo representa una temporada y podría variar si incluyera información que cubriera algunos años.

Palabras clave: SREG, GGE, MET, interacción, biomasa, *Brassica napus*.

LITERATURE CITED

- AFIPA. 2009-2010. Manual fitosanitario, Asociación Nacional de Fabricantes e Importadores de Productos Fitosanitarios Agrícolas A.G., Santiago, Chile.
- Crossa, J. 1990. Statistical analyses of multilocation trials. Advances in Agronomy 44:55-85.
- Crossa, J., P.N. Fox, W.H. Pfeiffer, S. Rajaram, and H.G. Gauch Jr. 1991. AMMI adjustment for statistical analysis of an international wheat yield trial. Theoretical and Applied Genetics 81:27-37.
- Diepenbrock, W. 2000. Yield analysis of winter oildseed rape (*Brassica napus* L.): A review. Field Crops Research 67:35-49.
- FAOSTAT. 2010. *Brassica napus*. Available at http:// faostat.fao.org (accessed 30 November 2010).
- Franco, J., J. Crossa, S. Taba, and H. Shands. 2003. A multivariate method for classifying cultivars and studying group x environment x trait interaction. Crop Science 43:1249-1258.
- Gabriel, K.R. 1971. The biplot-graphical display of matrices with applications to principal components analysis. Biometrika 58:453-467.
- Gauch, H.G. 2006. Statistical analysis of yield trials by AMMI and GGE. Crop Science 46:1488-1500.
- Kaya, Y., M. Ankura, and S. Taner. 2006. GGE-Biplot analysis of multi-environment yield trials in bread wheat. Turkish Journal of Agriculture and Forestry 30:325-337.
- Kempton, R.A. 1984. The use of biplots in interpreting variety by environment interactions. Journal of Agricultural Science 103:123-135.
- Long, Y., J. Shi, D. Qiu, C. Zhang, J. Wang, J. Hou, J. Zhao, *et al.* 2007. Flowering time quantitative trait loci analysis of oilseed Brassica in multiple environments and genome wide alignment with Arabidopsis. Genetics 177:2433-2444.
- Ma, B.L., W. Yan, L.M. Dwyer, J. Frégeau-Reid, H.D. Voldeng, Y. Dion, and H. Nass. 2004. Graphic analysis of genotype, environment, nitrogen fertilizer, and their interactions on spring wheat yield. Agronomy Journal 96:169-180.
- Major, D.J., J.B. Bole, and W.A. Charnetski. 1978. Distribution of photosynthates after ¹⁴CO₂ assimilation by stem, leaves, and pods of rape plants. Canadian Journal of Plant Science 58:783-787.
- Nassimi, A., W. Raziuddin, and A. Naushad. 2006. Heterotic studies for yield associated traits in *Brassica napus* L. using 8 x 8 diallel crosses. Pakistan Journal of Biological Science 9:2132-2136.

- Navabi, A., R.-C. Yang, J. Helm, and D.M. Spaner. 2006. Can spring wheat-growing mega-environments in the northern Great Plains be dissected for representative locations or niche adapted genotypes? Crop Science 46:1107-1116.
- Nielsen, D.C. 1996. Potential of canola as a dryland crop in northeastern Colorado. p. 281-287. *In* J. Janick (ed.) Progress in new crops. ASHS Press, Alexandria, Virginia, USA.
- ODEPA. 2009. Raps canola. Oficina de Estudios y Políticas Agrarias (ODEPA), Ministerio de Agricultura, Santiago, Chile. Available at http://www. odepa.gob.cl (accessed 2 June 2009).
- Ortegón, A.S., F.A. Díaz, y Q.J González. 2007. Cultivares de canola y su interacción con el ambiente y el método de siembra. Rev. Instituto Nacional de Investigaciones Forestales, Agrícolas y Pecuarias. Campo Experimental Río Bravo, UNIFAP. Universidad y Ciencia 23:21-28.
- Ortegón. A.S., A.D. Franco, y A. Ramírez de León. 2006. Rendimiento y calidad de semilla de variedades e híbridos de canola en el Norte de Tamaulipas, México. Revista Fitotecnia Mexicana 29:181-186.
- Radoev, M., H.C. Becker, and W. Ecke. 2008. Genetic analysis of heterosis for yield and yield components in rapeseed (*Brassica napus* L.) by quantitative trait locus mapping. Genetics 179:1547-1558.
- Rodríguez, J., J. Sahagún, H. Villaseñor, J. Molina, y A. Martínez. 2002. Estabilidad de siete variedades comerciales de trigo (*Triticum aestivum* L.) de Temporal. Revista Fitotecnia Mexicana 25:143-151.
- Roozeboom, K.L., T.W. Schapaugh, M.R. Tuinstra, R.L. Vanderlip, and G. Milliken. 2008. Testing wheat in variable environments: Genotype, environment, interaction effects, and grouping test locations. Crop Science 48:317-330.
- Sabaghnia, N., H. Dehghani, and S. Sabaghpour. 2008. Graphic analysis of genotype by environment interaction for lentil yield in Iran. Agronomy Journal 100:760-764.
- SAS Institute. 2007. SAS/STAT® User's guide. Version 8.1. 8th ed. Vol. 1. SAS Institute, Cary, North Carolina, USA.
- Thomas, P. 2003. The growers manual. Canola Council of Canada. Winnipeg, Canada. Available at http://www. canola-council.org/CouncilPublications (accessed 26 September 2009).
- Vargas, M., y J. Crossa. 2000. El análisis AMMI y la gráfica del biplot en SAS. CIMMYT. México D.F., México. Available at http://www.cimmyt.org/ biometrics (accessed July 2008).

- Yan, W. 2001. GGE biplot-A windows application for graphical analysis of multi environment trial data and other types of two-way data. Agronomy Journal 93:1111-1118.
- Yan, W. 2007. GGE biplot. Version 6. Available at http:// www.ggebiplot.com (accessed 26 October 2009).
- Yan, W., L.A. Hunt, Q. Sheng, and Z. Szlavnics. 2000. Cultivar evaluation and mega-environment investigation based on the GGE biplot. Crop Science 40:597-605.
- Yan, W., M. Kang, B. Ma, S. Woods, and P. Cornelius. 2007. GGE biplot vs. AMMI analysis of genotype-byenvironment data. Crop Science 47:643-653.

- Yan, W., and I. Rajcan. 2002. Biplot evaluation of test sites and trait relations of soybean in Ontario. Crop Science 42:11-20.
- Yang, R., J. Crossa, P. Cornelius, and J. Bugueño. 2009. Biplot analysis of genotype x environment interaction: Proceed with caution. Crop Science 49:1564-1576.
- Zobel, R.W., M.J. Wright, and H.G. Gauch Jr. 1988. Statistical analysis of a yield trial. Agronomy Journal 80:388-393.