

# Yield stability and relationships among stability parameters in soybean genotypes across years

Anderson S. Milioli<sup>1</sup>, Andrei D. Zdziarski<sup>1</sup>, Leomar G. Woyann<sup>1</sup>, Rodnei dos Santos<sup>1</sup>, Ana C. Rosa<sup>1</sup>, Alana Madureira<sup>1</sup>, and Giovani Benin<sup>1\*</sup>

<sup>1</sup>Universidade Tecnológica Federal do Paraná – UTFPR, Campus Pato Branco. Via do Conhecimento, km 01, CEP 85503-390, Pato Branco - Paraná, Brasil. \*Corresponding author (benin@utfpr.edu.br).

Received: 28 January 2018; Accepted: 11 May 2018; doi:10.4067/S0718-58392018000200299

# ABSTRACT

The search for productive and stable genotypes is the main goal of breeding programs. The Genotype  $\times$  Environment interaction strongly influences genotype performance, and makes the selection of new cultivars difficult. One way to take advantage of this interaction is to identify genotypes with high grain yield (GY) and stability in different environments. The objective of this study was to evaluate the consistency of correlation between GY and stability evaluation methods in multi-environment trials and identify which methods are more suitable for selecting genotypes. GY data from 11 soybean (Glycine max [L.] Merr.) cultivars conducted in Value for Cultivation and Use trials in 10 locations in Paraná and Mato Grosso do Sul states, Brazil, in the 2013-2014, 2014-2015, and 2015-2016 crop seasons. A randomized complete block design with three replicates was used. Seven methods were applied to evaluate stability, and Spearman's correlation coefficient was used to compare methods. Positive associations were observed between GY and the harmonic mean of genotypic values (HMGV) across environments and genotype main effect + Genotype × Environment interaction effect by ideal genotype (GGE IG) methodologies, and between GY and the Lin and Binns method modified by Carneiro for general and unfavorable environments. The Eberhart and Russell, additive main effects and multiplicative interaction (AMMI1), and GGE for stability (GGE STA) methods presented no positive associations with GY in any year. Positive associations were found between the Wricke, AMMI1, and Eberhart and Russell methods because they were related to the static stability concept. The HMGV and GGE IG methods can be used together because they are consistently associated with GY and based on the dynamic stability concept.

Key words: Consistency between years, dynamic and static stability, Genotype × Environment interaction, Glycine max.

#### INTRODUCTION

Soybean (*Glycine max* [L.] Merr.) is grown in an extensive cultivation area in Brazil, and is the main commodity of the national agricultural sector. World estimated production for the 2016-2017 crop season was approximately 336 million tons, and Brazil was responsible for over 107 million tons (Conab, 2017).

The search for genotypes with high grain yield and stability is the key aspect of plant breeding programs, which aim to obtain genotypes with high performance for different cultivation regions. The Genotype × Environment (GE) interaction strongly influences genotype performance, making new cultivar selection difficult. One way to take advantage of this interaction is to identify genotypes with high production capacity and phenotypic stability in different environments (Bornhofen et al., 2017).

Identifying genotypes with high grain yield and stability is the main goal of plant breeding programs. The literature describes numerous methods to study the stability and productive performance of genotypes. These methods differ in the

evaluated parameters and statistics. Studies comparing different methods have already been described for different crops, such as wheat (Bornhofen et al., 2017), maize (Cargnelutti Filho et al., 2009), common beans (Pereira et al., 2009), and soybean (Polizel et al., 2013).

Among the main methods used to study phenotypic stability of genotypes are univariate parametric models (Wricke, 1965; Eberhart and Russell, 1966), multivariate parametric models (Zobel et al., 1988; Yan, 2001), non-parametric models (Lin and Binns, 1988), and mixed models (Resende, 2006). The ability to explain the sum of squares of the GE interaction is the main difference among methods (Bornhofen et al., 2017).

Phenotypic stability can be divided into two principal types, that is, stability in the biological and agronomic sense. Stability in the biological (static) sense refers to the ability of genotypes to maintain constant production in different environments, with low variation between them, that is, genotypes exhibit "homeostasis". Stability in the agronomic (dynamic) sense indicates that the genotype positively responds to improvements in edaphoclimatic conditions of the environment and can perform above the mean in different locations (Sabaghnia et al., 2015). This behavior is of interest to plant breeders and farmers. It is important for the plant breeder to adopt methods in which genotype stability is associated with high grain yield average. Among the different currently available methods, the genotype main effect + genotype x environment (GGE) biplot enables simultaneous analysis of genotypes by considering high grain yield average and stability. The combination of these two concepts is known as the "ideal genotype" (Yan et al., 2007; Yan, 2016).

To compare the results obtained between different methods, many authors have used Spearman's correlation. This comparison methodology is based on ranks. However, it is not known if the methods provide a consistent result regarding the correlation between the different years under study. The new contribution of the present study is the evaluation of the consistency of Spearman's rank correlation between different years using multivariate methods, such as genotype main effect + genotype × environment (GGE) interaction effect, additive main effects and multiplicative interaction (AMMI1), and mixed models. Therefore, the objective of this study was to evaluate the consistency of association between grain yield and stability methods in multi-environment trials and identify which of the evaluated methods is more suitable for selecting superior genotypes.

## MATERIALS AND METHODS

Soybean grain yield (GY) data were obtained from trials of Value for Cultivation and Use (VCU) at 10 locations in the 2013-2014, 2014-2015, and 2015-2016 crop seasons. The locations are in the soybean macro-region 2, situated in the states of Paraná and Mato Grosso do Sul, Brazil. In Paraná, trials were conducted at Bela Vista do Paraíso (23°00' S, 51°11' W), Cafelândia (24°37' S, 53°19' W), Floresta (23°36' S, 52°05' W), Palotina (24°17' S, 53°50' W), Rolândia (23°19' S, 51°22' W), Santa Terezinha de Itaipu (25°27' S, 54°24' W), and Sertanópolis (23°04' S, 51°02' W). In Mato Grosso do Sul, trials were conducted at Dourados (22°13' S, 54°49' W), Maracaju (21°37' S, 55°10' W), and Naviraí (23°04' S, 54°11' W).

Eleven soybean cultivars were evaluated (coded as C1 to C11): 5958RSF IPRO (C1), 6563RSF IPRO (C2), NA 5909 RG (C3), BMX Potência RR (C4), DM 5.9i (C5), NK 7059 RR (C6), 6160RSF IPRO (C7), 6458RSF IPRO (C8), 68I70RSF IPRO (C9), 61I59RSF IPRO (C10), and 63I64RSF IPRO (C11). Trials were carried out in a randomized complete block design with three replicates. Each experimental plot consisted of four rows, 5 m long, and 0.5 m row spacing. The two central rows were mechanically harvested from each plot (5 m<sup>2</sup>). Sowing density was standardized at 350 000 seeds ha<sup>-1</sup> for all cultivars. Crop management followed the technical recommendations for soybean cultivation. Grain moisture was adjusted to 13% and expressed in kg ha<sup>-1</sup>.

Data were tested for normality according to the Lilliefors test. Joint and individual ANOVA were performed each year. The significance of genotype (G), environment (E), and G×E (GE) interaction effects were determined by the F test. Selective accuracy was calculated according to the method described by Resende and Duarte (2007). Genotype performance for the group of environments was evaluated each year by the Scott-Knott method at 5% error probability. The method described by Annicchiarico (1992) was used to divide the environments as favorable and unfavorable in 2013, 2014, and 2015. After this division, ANOVA was performed for the environments classified as favorable and unfavorable. A new group of means was then performed by the Scott-Knott method. This procedure aimed to identify differences in cultivar grouping for the group of environments and favorable and unfavorable environments.

In order to evaluate stability, the following methods were tested: Wricke (1965) ( $\omega$ i), Eberhart and Russel (1966) (E-R), Lin and Binns modified by Carneiro (1998) (L-B/C), additive main effects and multiplicative interaction (AMMI1) (Zobel et al., 1988), and the harmonic mean of the genotypic values (HMGV) across environments via mixed models by restricted maximum likelihood (REML)/best linear unbiased prediction (BLUP), model 54 (Resende, 2006). In addition, for multivariate analysis via GGE, the biplot graph and output (Log) values of the analyses were used to identify the most stable genotypes. The GGE biplot (Yan, 2001), Selegen (Resende, 2006), and Genes (Cruz, 2013) software were used to perform the analyses.

Spearman's rank correlation coefficient was used to compare the described methodologies. Cultivars were ranked by considering stability and productive performance. Thus, the genotypes were first ordered by GY based on mean performance for the complete set of environments; the best genotype performance was allotted number 1 and the worst number 11.

For the  $\omega$  parameter, genotype ranking was carried out by the ecovalence index. In this method, when the modular ecovalence of the genotype is lower, stability is higher. The genotype that exhibited the lowest modular ecovalence value was allotted number 1 and the genotype with the highest value was number 11 in the ranking.

In the methodology described by E-R, genotypes were arranged according to the deviation of the regression. Genotypes that showed no significant deviation of the regression ( $S^2d = 0$ ) were classified as stable. In this group, the genotype with the lowest deviation of the regression in the module was ranked as 1 and up to the g<sup>th</sup> deviation of the regression. Then, genotypes with significant deviation of the regression at 5% and 1% were ranked following the same procedure.

The L-B/C method is based on the  $P_i$  parameter where the most stable genotype has the lowest  $P_i$  value. The modification proposed by Carneiro (1998) enables the identification of stable genotypes in favorable, unfavorable, and general environments. Ranking genotypes by  $P_i$  (g),  $P_i$  (f), and  $P_i$  (u) parameters was carried out for general, favorable, and unfavorable environments, respectively. The genotype with the lowest Pi value was classified as the most stable and ranked as 1. The process was repeated for the 11 genotypes and three parameters.

The HMGV parameter was used for classification by the mixed model method because it estimates stability. In the multivariate methods, AMMI1 and GGE for stability (GGE STA), genotype stability was evaluated by the graphs and log values of the GGE biplot software. In these methods, vector projection in relation to the AEC-axis, regardless of the direction, represents stability, and when the vector is smaller, stability is higher. In the ideal genotype analysis (GGE IG), a value of 1.5 was used as weight for mean GY in relation to stability.

Spearman's rank correlation coefficient was used to compare the methods to measure stability. Consistency of the methods between years was evaluated by the significance of the associations and the coexistence of similar correlations in the 3 years. When results were similar in the correlations for the 3 years, the correlated methods were accepted as consistent, which were significant or nonsignificant by the t test for n = 11. The magnitude of the coefficients of Spearman's correlation was according to the description by Monteiro et al. (2015) as follows: null magnitude (0.0 to 0.20), weak magnitude (0.21 to 0.40), average magnitude (0.41 to 0.70), strong magnitude (0.71 to 0.90), and extremely strong magnitude (0.91 to 1.00).

#### **RESULTS AND DISCUSSION**

The joint ANOVA for locations in each year indicated that the sources of variation of G, E, and GEI were significant (p < 0.01) for all the evaluated years (Table 1). This indicates that genotypes exhibited different behavior in the different environments, and that there was a change in the magnitude of the responses for GY due to environmental variation. Similar results were observed by Cargnelutti Filho et al. (2007). The GY mean of the trials was 3529.34 kg ha<sup>-1</sup> in 2013, 3333.03 kg ha<sup>-1</sup> in 2014, and 3486.42 kg ha<sup>-1</sup> in 2015. The coefficient of variation (CV) was 7.96%, 9.74%, and 10.23%, and selective accuracy was 0.70, 0.86, and 0.87 in 2013, 2014, and 2015, respectively. These results indicate high experimental precision, providing security and reliability for selecting superior genotypes.

The GY means for the cultivars in the 3 years and 10 locations are shown in Table 2. The 11 cultivars were present in the 10 locations for the 3 years, and the inferences on stability were made for the same cultivars and locations in the different years. The difference between the cultivars was nonsignificant for the 10 locations as a group and for each of the three evaluated years. This was probably due to the complex interaction between cultivars and the experimental locations that changed the genotype ranking between locations evaluated within the same year (Costa et al., 2015). The genetic

	ANOVA	(2013)				
Source	DF	MS	F-test			
Blocks/E	20	103727.50	-			
Genotypes (G)	10	1293605.65	16.4**			
Environments (E)	9	80841312.84	779.36**			
G×E	90	655677.82	8.31**			
Error	200	-				
Mean, kg ha <sup>-1</sup> = $3529.34$		CV (%) = 7.96				
> MS/ $<$ MS ratio = 6.07						
	ANOVA	(2014)				
Source	DF	MS	F-test			
Blocks/E	20	181539.91	-			
Genotypes (G)	10	16.77**				
Environments (E)	9	71.39**				
G×E	90	4.45**				
Error	200	105292.14	-			
Mean, kg ha <sup>-1</sup> = $3333.03$		CV (%) = 9.74				
> MS/ $<$ MS ratio = 6.84	Selective ac					
	ANOVA	(2015)				
Source	DF	MS	F-test			
Blocks/E	20	107415.12	-			
Genotypes (G)	10	1601149.66	12.59**			
Environments (E)	9	5616124.46	52.28**			
G×E	90	3.02**				
Error	200	-				
Mean, kg ha <sup>-1</sup> = $3486.42$		CV (%) = 10.23				
> MS/ $<$ MS ratio = 6.99	Selective accuracy $= 0.87$					

Table 1. Joint analysis of variance (ANOVA) for soybean grain yield (GY) of 11 cultivars at 10 locations in 2013, 2014, and 2015.

\*\*Significant at 0.01 probability level according to F-test.

MS: Mean square; CV: coefficient of variation; DF: degree of freedom;

G: Genotype; E: Environment; G × E: Genotype-Environment interaction.

correlation between performances in the evaluated environments (rgloc) was 0.10, 0.26, and 0.32 in 2013, 2014, and 2015, respectively (data not shown), indicating extremely complex GE interaction. Values of rgloc that are close to 0.50 indicate complex GE interaction with alterations in the ranking of cultivars between environments (Carvalho et al., 2016). This shows the need for statistical methods that enable the identification of superior genotypes with high GY and stability.

The absence of significant difference between genotypes for the group of 10 locations indicates the need to discriminate environments. The method by Annicchiarico (1992) allows classifying environments as favorable and unfavorable based on grain yield at each location (Table 3). After designating environments as favorable and unfavorable, a significant difference was observed between years (Table 2). Among the environments classified as favorable, differentiation between cultivars was found only in 2013 and cultivars C2, C7, C8, and C10 showed superior performance. Regarding unfavorable environments, both the 2013 and 2014 seasons exhibited significant differences between cultivars. In 2013, cultivars C2, C4, C6, C8, C9, C10, and C11 were superior, while cultivar C9 had superior performance in 2014. In 2015, classifying environments as favorable and unfavorable was ineffective to identify superior cultivars for GY.

The comparison of statistical methods is shown in Table 4. Spearman's correlation coefficient varied from -0.89 to 1.00. This result indicates different consistency categories between the parameters of cultivar classification. Significant positive associations were observed between GY and stability by the L-B/C methodology (Table 4). The magnitude of the associations of the L-B/C method with GY for general environments (P<sub>i</sub>) and unfavorable environments (P<sub>i</sub>(u)) were 0.65 and 0.83, 0.91 and 0.77, and 0.99 and 0.92 in 2013, 2014, and 2015, respectively. For the favorable environments (P<sub>i</sub>(f)), the correlation with GY was significant in 2014 (0.85) and 2015 (0.97). In 2013, the correlation between GY and P<sub>i</sub>(f) was nonsignificant (0.27). Therefore, the P<sub>i</sub>(f) parameter was not consistent between years. The evaluation of productive stability by non-parametric methods, such as the Lin and Binns (1988) method, is advantageous since it combines GY and stability (Zali et al., 2011). However, the lack of consistency between years can generate unreliable information and is associated with GY only in some years.

							Parameters							
Cultivar	Year	GY	F	U	Rank	HMGV	GGE STA	AMMI1	ωi	Pi	Pi(f)	Pi(u)	S <sup>2</sup> d	GGE IG
			- kg ha <sup>-1</sup>											
C1	2013	3380a	5170b	2186b	9	8	6	7	6	9	7	8	7	9
	2014	3259a	3722a	2566c	8	8	3	4	1	8	5	8	1	8
	2015	3281a	3602a	2800a	8	8	3	8	6	8	8	9	6	8
C2	2013	3793a	5611a	2581a	2	1	1	1	3	1	1	2	4	2
	2014	3414a	3800a	2836c	5	4	6	1	7	3	3	3	7	2
	2015	3539a	3769a	3193a	6	6	9	1	7	6	7	5	7	6
C3	2013	3333a	5209b	2083b	10	9	5	3	2	6	5	9	1	8
	2014	3176a	3566a	2592c	9	9	2	3	2	9	9	7	2	9
	2015	3655a	3966a	3188a	4	4	2	4	3	3	3	6	3	3
C4	2013	3487a	4925b	2529a	7	5	10	10	8	7	9	6	8	3
	2014	3511a	3780a	3107b	2	2	9	10	9	2	4	2	9	3
	2015	3727a	4014a	3297a	2	2	5	11	11	2	2	4	11	1
C5	2013	3157a	4939b	1969b	11	11	7	8	5	11	8	11	6	11
	2014	2914a	3413a	2165d	11	11	1	5	3	10	10	11	4	11
	2015	3144a	3435a	2706a	11	11	7	6	5	11	11	10	5	10
C6	2013	3508a	5165b	2403a	6	7	3	2	4	4	6	7	3	5
	2014	3270a	3636a	2721c	7	7	8	6	8	7	8	6	8	7
	2015	3274a	3509a	2921a	9	9	4	9	8	9	10	8	8	9
C7	2013	3407a	5508a	2006b	8	10	9	9	7	5	2	10	5	7
	2014	3014a	3457a	2349d	10	10	10	11	10	11	11	10	10	10
	2015	3178a	3502a	2692a	10	10	10	10	10	10	9	11	10	11
C8	2013	3717a	5467a	2550a	3	3	4	4	1	2	3	4	2	4
	2014	3428a	3830a	2824c	4	3	5	7	4	5	7	5	5	6
	2015	3480a	3803a	2997a	7	7	1	2	1	7	6	7	2	7
C9	2013	3535a	5015b	2548a	5	6	11	11	11	10	11	3	11	6
	2014	3776a	3847a	3669a	1	1	11	11	11	1	2	1	11	1
	2015	3694a	3937a	3328a	3	3	11	5	9	4	4	2	9	4
C10	2013	3822a	5672a	2588a	1	4	8	5	9	3	4	5	9	1
	2014	3395a	3753a	2858c	6	6	7	2	5	4	6	4	6	4
	2015	3592a	3795a	3288a	5	5	8	7	4	5	5	3	4	5
C11	2013	3686a	5248b	2644a	4	2	2	6	10	8	10	1	10	10
	2014	3506a	4072a	2657c	3	5	4	8	6	6	1	9	3	5
	2015	3787a	4025a	3430a	1	1	6	3	2	1	1	1	1	2

Table 2. Ranking of 11 soybean cultivars evaluated at 10 locations during 3 years (2013, 2014, and 2015), as related to grain yield (GY) in all environments, in favorable (F) and unfavorable (U) environments, and parameters of stability methods.

<sup>1</sup>Means followed by the same letter in the same year do not differ according to the Scott-Knott test (p = 0.05).

C: Cultivar; GY: grain yield; Parameters to evaluate adaptability and stability:  $\omega$ : ecovalence (Wricke, 1965); S<sup>2</sup>d: Eberhart and Russell (1966); Pi, Pi(f) and Pi(u): Lin and Binns (1988) method modified by Carneiro (1998); HMGV: stability by mixed models (REML/BLUP); AMMI1: additive main effects and multiplicative interaction, first principal component of the AMMI1 analysis; GGE: genotype main effect + genotype x environment interaction, stability by GGE biplot analysis; i, (f), and (u): performance in general, favorable environments, and unfavorable environments, respectively.

Cultivars: C1: 5958RSF IPRO, C2: 6563RSF IPRO, C3: NA 5909 RG, C4: BMX Potência RR, C5: DM 5.9i, C6: NK 7059 RR, C7: 6160RSF IPRO, C8: 6458RSF IPRO, C9: 68170RSF IPRO, C10: 61159RSF IPRO, and C11: 63164RSF IPRO.

Table 3. Classification (Class) of locations as favorable (F) or unfavorable (U) according to method by Annicchiarico (1992) for soybean grain yield in 2013, 2014, and 2015.

,							
	20	013	2	014	2015		
Locations	Class	Mean	Class	Mean	Class	Mean	
		kg ha <sup>-1</sup>		kg ha <sup>-1</sup>		kg ha <sup>-1</sup>	
Bela Vista do Paraíso	U	2598	U	2933	F	3544	
Cafelândia	F	4687	U	3010	U	2973	
Dourados	U	2328	F	3352	F	3764	
Floresta	U	2009	U	2841	F	3667	
Maracaju	F	4955	F	3405	U	3183	
Naviraí	U	2291	U	2249	F	3567	
Palotina	F	6126	F	3644	F	3714	
Rolândia	U	2370	F	3408	U	3068	
Santa Terezinha de Itaipu	F	5297	F	4036	F	4302	
Sertanópolis	U	1931	F	4452	U	3082	

Parameter	Year	S <sup>2</sup> d	Pi	Pi(f)	Pi(u)	ωi	AMMI1	GGE STA	HMGV	GGE IG
Pi	2013	0.52 <sup>ns</sup>								
	2014	-0.43 <sup>ns</sup>								
	2015	0.21 <sup>ns</sup>								
Pi(f)	2013	0.66**	0.83**							
	2014	-0.05 <sup>ns</sup>	0.78**							
	2015	0.24 <sup>ns</sup>	0.98**							
Pi(u)	2013	0.38 <sup>ns</sup>	0.31 <sup>ns</sup>	-0.13 <sup>ns</sup>						
	2014	-0.51 <sup>ns</sup>	0.93**	0.56 <sup>ns</sup>						
	2015	0.22 <sup>ns</sup>	0.88**	0.84**						
ωi	2013	0.95**	0.49 <sup>ns</sup>	0.63**	-0.28 <sup>ns</sup>					
	2014	0.95**	-0.44 <sup>ns</sup>	-0.24 <sup>ns</sup>	-0.45 <sup>ns</sup>					
	2015	0.99**	0.15 <sup>ns</sup>	0.19 <sup>ns</sup>	0.16 <sup>ns</sup>					
AMMI1	2013	0.66**	0.71**	0.60 <sup>ns</sup>	0.19 <sup>ns</sup>	0.69**				
	2014	0.59 <sup>ns</sup>	-0.09 <sup>ns</sup>	-0.07 <sup>ns</sup>	-0.04 <sup>ns</sup>	0.67**				
	2015	0.67**	0.25 <sup>ns</sup>	0.23 <sup>ns</sup>	0.35 <sup>ns</sup>	0.68**				
GGE STA	2013	0.47 <sup>ns</sup>	0.46 <sup>ns</sup>	0.35 <sup>ns</sup>	0.35 <sup>ns</sup>	0.55 <sup>ns</sup>	0.85**			
	2014	0.93**	-0.49 <sup>ns</sup>	-0.20 <sup>ns</sup>	-0.57 <sup>ns</sup>	0.92**	0.60*			
	2015	0.47 <sup>ns</sup>	0.07 <sup>ns</sup>	0.09 <sup>ns</sup>	-0.19 <sup>ns</sup>	0.52 <sup>ns</sup>	0.08 <sup>ns</sup>			
HMGV	2013	-0.20 <sup>ns</sup>	0.54 <sup>ns</sup>	0.11 <sup>ns</sup>	0.94**	-0.08 <sup>ns</sup>	0.36 <sup>ns</sup>	0.45 <sup>ns</sup>		
	2014	-0.43 <sup>ns</sup>	0.95**	0.77**	0.86**	-0.47 <sup>ns</sup>	-0.29 <sup>ns</sup>	-0.52 <sup>ns</sup>		
	2015	0.15 <sup>ns</sup>	0.99**	0.97**	0.92**	0.10 <sup>ns</sup>	0.24 <sup>ns</sup>	-0.01 <sup>ns</sup>		
GGE IG	2013	0.05 <sup>ns</sup>	0.75**	0.45 <sup>ns</sup>	0.41 <sup>ns</sup>	0.06 <sup>ns</sup>	0.28 <sup>ns</sup>	-0.04 <sup>ns</sup>	0.57 <sup>ns</sup>	
	2014	-0.44 <sup>ns</sup>	0.97**	0.84**	-0.89**	-0.50 <sup>ns</sup>	-0.07 <sup>ns</sup>	-0.54 <sup>ns</sup>	0.92**	
	2015	0.16 <sup>ns</sup>	0.98**	0.95**	0.86**	0.12 <sup>ns</sup>	0.20 <sup>ns</sup>	0.11 <sup>ns</sup>	0.97**	
GY (kg ha <sup>-1</sup> )	2013	-0.23 <sup>ns</sup>	0.65*	0.27 <sup>ns</sup>	0.83**	-0.16 <sup>ns</sup>	0.34 <sup>ns</sup>	0.27 <sup>ns</sup>	0.88**	0.73*
	2014	-0.37 <sup>ns</sup>	0.91**	0.85**	0.77**	-0.48 <sup>ns</sup>	-0.36 <sup>ns</sup>	-0.49 <sup>ns</sup>	0.97**	0.90**
	2015	0.15 <sup>ns</sup>	0.99**	0.97**	0.92**	0.10 <sup>ns</sup>	0.24 <sup>ns</sup>	-0.01 <sup>ns</sup>	1.00**	0.97**

Table 4. Repeatability of Spearman's correlation coefficients between evaluation methods of grain yield and stability concepts using rankings of 11 soybean cultivars evaluated in 2013, 2014, and 2015.

\*, \*\*Significant at the 0.05 and 0.01 probability levels according to Student's t-test, respectively; <sup>ns</sup> nonsignificant according to Student's t-test at 0.05 probability level. GY: grain yield; Stability parameters:  $\omega$ i (Wricke, 1965); S<sup>2</sup>d (Eberhart and Russell, 1966); Pi, Pi(f), and Pi(u) for stability of environments in general, favorable environments, or unfavorable environments according to the method by Lin and Binns modified by Carneiro (1998); additive main effects and multiplicative interaction – AMMI1 (Zobel et al., 1988); additive main effect of genotype plus multiplicative interaction of genotype main effect + genotype × environment interaction (GGE) for stability (GGE STA) and ideal genotype (GGE IG) (Yan, 2001); mixed models – REML/BLUP, model 54 (Resende, 2006); HMGV: stability obtained by harmonic mean of the genotypic values across environments .

Correlation coefficients between GY and the HMGV parameter were 0.88, 0.97, and 1.00 in 2013, 2014, and 2015, respectively. The observed correlations in all 3 years indicate that these methods enable the identification of stable genotypes with high GY, that is, they are associated with the dynamic concept of stability (Annicchiarico, 2002). The HMGV method classifies genotypes by their estimated genetic values and also by stability. Therefore, this method is different because it evaluates genotypes in terms of estimated genetic values by best linear unbiased prediction (BLUP) and restricted maximum likelihood (REML) (Resende and Duarte, 2007). On the other hand, GY was not positively associated with the E-R,  $\omega$ i, AMMI1, and GGE STA methodologies in any year. These results indicate that genotypes with high stability are not always the most productive; results concur with those reported by Bornhofen et al. (2017) when evaluating wheat genotypes.

Significant positive associations were found between the  $\omega$ i and AMMI1 (0.69, 0.67, and 0.68) and E-R (0.95, 0.95, and 0.99) methodologies in 2013, 2014, and 2015, respectively. Other authors have also found positive associations between these methods (Paula et al., 2014; Bornhofen et al., 2017). This occurs because these methods are associated with the concept of static stability, which is obtained independently of genotype mean yield (Annicchiarico, 2002). The selection exclusively made by these methods can result in selecting stable but not necessarily productive genotypes.

The E-R (S<sup>2</sup>d) methodology showed a significant association with the  $P_i(f)$  parameter (0.66) from the L-B/C method only in 2013. However, Dehghani et al. (2008) report a negative association (-0.55) between the L-B/C and E-R methods. In addition, an association was observed between L-B/C and the  $\omega i$  and AMMI1 methodologies only in 2013, and the association with the GGE STA parameter was absent. There is therefore no consistency between years, indicating differences between methods because they are based on different concepts of stability. Representatives of both groups should be used simultaneously in practice to consider the different concepts of stability and enable more reliable selection and recommendation of genotypes. Significant positive associations between different methods indicate that genotypes are similarly classified as to stability. Methods may be supplying redundant information, and only one would be sufficient to select the best genotypes (Cargnelutti Filho et al., 2009). However, even with strong association between methods, genotype ranking can be different between methodologies. This is the case, for example, of the association between GY and HMGV in 2013, that is, the correlation is high (0.88) but the methods differ as to the best cultivar; cultivar C2 was superior in HMGV, while cultivar C10 was superior for GY. In the HMGV method, cultivar C10 was only the fourth best genotype. Moreover, the HMGV method shows genotypic variation, whereas GY shows phenotypic variation. It is therefore necessary to use two or more methodologies to ensure more efficient selection.

The methods based on multivariate statistics make it possible to visually evaluate genotype behavior and more clearly identify genotypes with characteristics of interest. The GGE method consists in analyzing biplot graphs to evaluate genotype stability and GY (Figures 1 A, B, and C). The most stable cultivar for 2013 was C2, followed by C11, C6, C8, C3, and C1. In 2014, cultivars C5 and C3 were the most stable. The most stable cultivars in 2015 were C8, C3, and C1. However, the stability analysis of this method did not show a significant association with GY in all 3 years. Considering only the stability parameter, the absence of association of the GGE STA method in 2013, 2014, and 2015, with associations of 0.27, -0.49, and -0.01, respectively, indicates that this method is consistent, but not correlated, with GY. Only using stability results is insufficient for selecting genotypes with superior GY performance. However, the analysis of mean versus stability on the GGE biplot allows selecting superior genotypes in terms of both yield and stability (Yan et al., 2007). When the genotype is more to the right of the biplot, it is more productive, whereas it is more stable when it is closer to the single arrow x-axis.

The GGE biplot software (Yan, 2001) allows identifying the best genotypes with "ideal genotype" analysis (GGE IG) by GY and stability statistics (Figures 1 D, E, and F). In this analysis, it was possible to attribute different weights to genotype GY in relation to stability, and this value varied from 1 to 4. Lower values indicated higher weight for GY, that is, productive performance has to be high for the position to be at the extreme right of the biplot. On the other hand, higher values (e.g., 4) allow easily identifying an ideal genotype because there is a greater demand for genotype stability. In this study, the genotype identified as ideal was the same as the one that had the most productive performance for the evaluated period because 1.5 GY/stability weight was used. Weights close to 4 can alter the identification of the ideal genotype, especially when highly productive genotypes are very unstable.

The GGE IG method was efficient for selecting stable and productive genotypes, and showed a significant association with GY in 2013, 2014, and 2015 (0.73, 0.90, and 0.97, respectively). These results validate the efficiency of the method for selecting superior genotypes and confirm its consistency between years.

The AMMI1 method (Figure 2) also simultaneously provides productive performance and stability in identifying superior cultivars. This method divides the main effects of genotype and environment and the effects of the GE interaction. It also combines ANOVA for the main effects of genotype and environment, represented on the x-axis, and the effect of the interaction of the principal components on the y-axis (Bose et al., 2014). Cultivars C10, C2, C8, and C11 were the most productive in 2013 and cultivar C2 was able to combine GY and stability among the evaluated environments. In 2014, cultivar C9 had superior productive performance, although it was the most unstable among the evaluated genotypes. In the same year, cultivar C2 exhibited high GY and stability. Cultivar C11 was the most productive in 2015, and C2 was the most stable, in 2013 and 2014. The AMMI1 method had no significant association with GGE IG in any of the years under study, which confirms the differences between the methods and the possibility of using them together as complementary analyses.

The use of stability and GY estimation methods are fundamental to identify highly productive and stable genotypes. Several parameters were positively correlated one with the other for the different years, thus indicating consistency in the results and helping to suggest more accurate methods for GY and stability analysis of the soybean crop. Due to the different concepts of stability, the breeder should prioritize using a group of methods that are related or unrelated to GY. Furthermore, the methods can be differentiated by using phenotypic data, or by estimating genetic values. The group of methods to be used can therefore consist of the HMGV and GGE IG methods since they are consistently associated with GY in different evaluated years. In this case, the HMGV method is emphasized because it uses genetic value prediction and the concept of dynamic stability, while GGE IG is pointed out for evaluating stability and GY together. Moreover, genotype selection by one of the  $\omega$ , AMMI1, or S<sup>2</sup>d parameters is recommended because they exhibit positive and consistent associations with each other in the different years under study. These methods use the concept of static stability

(not associated with yield). Among the different studied methods, the GGE IG method is highlighted. Despite its stability parameter to use the static concept, it allows verifying GY and the concept of ideal genotype, as well as the ease provided by graphical analysis.

Figure 1. Mean, stability, and ideal genotype for the 11 soybean cultivars evaluated in 2013, 2014, and 2015 at 10 locations. A, B, and C correspond to analysis of mean and stability in 2013, 2014, and 2015, respectively, and D, E, and F correspond to analysis of ideal genotype in 2013, 2014, and 2015, respectively.



The evaluated cultivars were C1: 5958RSF IPRO, C2: 6563RSF IPRO, C3: NA 5909 RG, C4: BMX Potência RR, C5: DM 5.9i, C6: NK 7059 RR, C7: 6160RSF IPRO, C8: 6458RSF IPRO, C9: 68I70RSF IPRO, C10: 61I59RSF IPRO, and C11: 63I64RSF IPRO.

CHILEAN JOURNAL OF AGRICULTURAL RESEARCH 78(2) APRIL-JUNE 2018

Figure 2. Additive main effects and multiplicative interaction 1 (AMMI1) analysis for the 11 soybean cultivars evaluated in 2013 (A), 2014 (B), and 2015 (C) at 10 locations.



The evaluated cultivars were C1: 5958RSF IPRO, C2: 6563RSF IPRO, C3: NA 5909 RG, C4: BMX Potência RR, C5: DM 5.9i, C6: NK 7059 RR, C7: 6160RSF IPRO, C8: 6458RSF IPRO, C9: 68170RSF IPRO, C10: 61159RSF IPRO, and C11: 63164RSF IPRO.

## CONCLUSIONS

The evaluated methodologies show different results regarding the consistency of associations between grain yield and stability evaluation methods across years.

The Lin and Binns method modified by Carneiro (L-B/C) exhibits positive associations with the Eberhart and Russel (E-R), Wricke ( $\omega$ i), and additive main effects and multiplicative interaction (AMMI1) methodologies, as well as with grain yield (GY), but it is not consistent across years and generates unreliable information.

The harmonic mean of the genotypic values (HMGV) across environments parameter and the genotype main effect + genotype × environment interaction effect by ideal genotype (GGE IG) method has consistent results and strong or

extremely strong associations with GY; it is the most adequate method to select productive and stable genotypes.

The  $\omega$ i methodology consistently and significantly correlates with AMMI1 and E-R, and it is recommended to prioritize one of these methodologies to avoid redundant results.

Genotype selection by the GGE biplot multivariate methodologies is valid when considering the concepts of mean, stability, and ideal genotype, not only the concept of stability in the biological sense without the interference of yield.

#### ACKNOWLEDGEMENTS

The authors thank the GDM Seeds Company and the breeders Marcos Norio Matsumoto, Nizio Fernando Giasson, and Jair Rogério Unfried for providing the dataset from the conducted experiments, and for contributions in accomplishing the present study. We also acknowledge the Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (Capes) for granting the master and doctoral scholarships.

#### REFERENCES

- Annicchiarico, P. 1992. Cultivar adaptation and recommendation from alfalfa trials in Northern Italy. Journal of Genetics and Breeding 46:269-278.
- Annicchiarico, P. 2002. Genotype × environment interaction: challenges and opportunities for plant breeding and cultivar recommendation. FAO Plant Production and Protection, Paper nr 174. FAO, Rome, Italy.
- Bornhofen, E., Benin, G., Storck, L., Woyann, L.G., Duarte, T., Stoco, M.G., et al. 2017. Statistical methods to study adaptability and stability of wheat genotypes. Bragantia 76(1):1-10. doi:10.1590/1678-4499.557.
- Bose, L.K., Jambhulkar, N.N., Pande, K., and Singh, O.N. 2014. Use of AMMI and other stability statistics in the simultaneous selection of rice genotypes for yield and stability under direct-seeded conditions. Chilean Journal of Agricultural Research 74:3-9. doi:10.4067/S0718-58392014000100001.
- Cargnelutti Filho, A., Perecin, D., Malheiros, E.B., e Guadagnin, J.P. 2007. Comparação de métodos de adaptabilidade e estabilidade relacionados à produtividade de grãos de cultivares de milho. Bragantia 66:571-578.
- Cargnelutti Filho, A., Storck, L., Riboldi, J., e Guadagnin, J.P. 2009. Associação entre métodos de adaptabilidade e estabilidade em milho. Ciência Rural 39(2):340-347.
- Carneiro, P.C.S. 1998. Novas metodologias de análise da adaptabilidade e estabilidade de comportamento. PhD thesis. Universidade Federal de Viçosa, Viçosa, Minas Gerais, Brasil.
- Carvalho, L.P., Farias, F.J.C., Morello, C.L., e Teodoro, P.E. 2016. Uso da metodologia REML/BLUP para seleção de genótipos de algodoeiro com maior adaptabilidade e estabilidade produtiva. Bragantia 75(3):314-321. doi:10.1590/1678-4499.275.
- Conab. 2017. Acompanhamento da safra brasileira de grãos. V. 4 SAFRA 2016/17- N. 6 Sexto levantamento. Março, 2017. Available at http://www.conab.gov.br (accessed 5 April 2017).
- Costa, A.F., Leal, N.R., Ventura, J.A., Gonçalves, L.S.A., Amaral Júnior, A.T., and Costa, H. 2015. Adaptability and stability of strawberry cultivars using a mixed model. Acta Scientiarum Agronomy 37:435-440. doi:10.4025/actasciagron.v37i4.18251.
- Cruz, C.D. 2013. GENES a software package for analysis in experimental statistics and quantitative genetics. Acta Scientiarum 35:271-276. doi:10.4025/actasciagron.v35i3.21251.
- Dehghani, H., Sabaghpour, S.H., and Sabaghnia, N. 2008. Genotype × environment interaction for grain yield of some lentil genotypes and relationship among univariate stability statistics. Spanish Journal of Agricultural Research 6:385-394. doi:10.5424/sjar/2008063-5292.
- Eberhart, S.A., and Russell, W.A. 1966. Stability parameters for comparing varieties. Crop Science 6:36-40. doi:10.2135/ cropsci1966.0011183X000600010011x.
- Lin, C.S., and Binns, M.R. 1988. A superiority measure of cultivar performance for cultivar × location data. Canadian Journal of Plant Science 68:193-198. doi:10.4141/cjps88-018.
- Monteiro, F.J.F., Peluzio, J.M., Afférri, F.S., Carvalho, E.V., e Santos, W.F. 2015. Correlação entre parâmetros de quatro metodologias de adaptabilidade e estabilidade em cultivares de soja em ambientes distintos. Revista de la Facultad de Agronomía, La Plata 114(2):143-147.
- Paula, T.M., Marinho, C.D., Souza, V., Barbosa, M.H.P., Peternelli, L.A., Kimbeng, C.A., et al. 2014. Relationships between methods of variety adaptability and stability in sugarcane. Genetics and Molecular Research 13(2):4216-4225. doi:10.4238/2014.June.9.7.
- Pereira, H.S., Melo, L.C., Peloso, M.J., Faria, L.C., Costa, J.G.C., Díaz, J.L., et al. 2009. Comparação de métodos de análise de adaptabilidade e estabilidade fenotípica em feijoeiro-comum. Pesquisa Agropecuária Brasileira 44(4):374-383.
- Polizel, A.C., Juliatti, F.C., Hamawaki, O.T., Hamawaki, R.L., e Guimarães, S.L. 2013. Adaptabilidade e estabilidade fenotípica de genótipos de soja no estado do Mato Grosso. Bioscience Journal 29(4):910-920.

- Resende, M.D.V. 2006. O software SELEGEN REML/BLUP. Embrapa Gado de Corte (Embrapa Gado de Corte Documentos), Campo Grande, Mato Grosso do Sul, Brasil.
- Resende, M.D.V., e Duarte, J.B. 2007. Precisão e controle de qualidade em experimentos de avaliação de cultivares. Pesquisa Agropecuária Tropical 37:182-194.
- Sabaghnia, N., Karimizadeh, R., and Mohammadi, M. 2015. Graphic analysis of yield stability in new improved lentil (*Lens culinaris* Medik.) genotypes using nonparametric statistics. Acta Agriculturae Slovenica 103(1):113-127. doi:10.14720/aas.2014.103.1.12.
- Wricke, G. 1965. Zur Berechnung der Ökovalenz bei Sommerweizen und Hafer. Zeitschrift für Pflanzenzüchtung 52:127-138.
- Yan, W. 2001. GGE biplot a windows application for graphical analysis of multienvironmental trial data and other types of two-way data. Agronomy Journal 93:1111-1118. doi:10.2134/agronj2001.9351111x.
- Yan, W. 2016. Analysis and handling of G × E in a practical breeding program. Crop Science 56:2106-2118. doi:10.2135/ cropsci2015.06.0336.
- Yan, W., Kang, M.S., Ma, B., Woods, S., and Cornelius, P.L. 2007. GGE biplot vs. AMMI analysis of genotype-by-environment data. Crop Science 47(2):643-653. doi:10.2135/cropsci2006.06.0374.
- Zali, H., Farshadfar, E., and Sabaghpour, S.H. 2011. Non-parametric analysis of phenotypic stability in chickpea (*Cicer arietinum* L.) genotypes in Iran. Crop Breeding Journal 1:85-96.
- Zobel, R.W., Wright, M.J., and Gauch, H.G. 1988. Statistical analysis of a yield trial. Agronomy Journal 80:388-393. doi:10.2134/agronj1988.00021962008000030002x.